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The *Si elegans* project at the interface of experimental and computational *Caenorhabditis elegans* neurobiology and behavior

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Abstract

Objective. In light of recent progress in mapping neural function to behavior, we briefly and selectively review past and present endeavors to reveal and reconstruct nervous system function in *Caenorhabditis elegans* through simulation. **Approach.** Rather than presenting an all-encompassing review on the mathematical modeling of *C. elegans*, this contribution collects snapshots of pathfinding key works and emerging technologies that recent single- and multi-center simulation initiatives are building on. We thereby point out a few general limitations and problems that these undertakings are faced with and discuss how these may be addressed and overcome. **Main results.** Lessons learned from past and current computational approaches to deciphering and reconstructing information flow in the *C. elegans* nervous system corroborate the need of refining neural response models and linking them to intra- and extra-environmental interactions to better reflect and understand the actual biological, biochemical and biophysical events that lead to behavior. Together with single-center research efforts, the *Si elegans* and OpenWorm projects aim at providing the required, in some cases complementary tools for different hardware architectures to support advancement into this direction. **Significance.** Despite its seeming simplicity, the nervous system of the hermaphroditic nematode *C. elegans* with just 302 neurons gives rise to a rich behavioral repertoire. Besides controlling vital functions (feeding, defecation, reproduction), it encodes different stimuli-induced as well as autonomous locomotion modalities (crawling, swimming and jumping). For this dichotomy between system simplicity and behavioral complexity, *C. elegans* has challenged neurobiologists and computational scientists alike. Understanding the underlying mechanisms that lead to a context-modulated functionality of individual neurons would not only advance our knowledge on nervous system function and its failure in pathological states, but have directly exploitable benefits for robotics and the engineering of brain-mimetic computational architectures that are orthogonal to current von-Neumann-type machines.

Keywords: *Caenorhabditis elegans*, brain-inspired computation, nervous system emulation, neuromorphic engineering

Introduction

Nervous systems are a special invention of nature for every multicellular organism 'on the move'. Living on a different timescale with other prey and defense mechanisms than hunt



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and escape, ‘rooted’ organisms such as plants, fungi or sponges process stimuli through a far less complex network of less specialized information processing cells compared to neurons in nervous systems (Desalle and Tattersall 2012). How complex does a nervous system need to become to let a multitude of adjustable stimulus-response actions (usually referred to as behavioral output) emerge? The nematode *Caenorhabditis elegans* (*C. elegans*) provides nature’s answer.

C. elegans, a tiny roundworm (*L*: 1 mm, \emptyset 80 μ m) with a life span of a few weeks, is among the five best characterized organisms in nature (Epstein and Shakes 1995). With less than 1% of its population being males, the nematode proliferates predominantly as a quasi-clone through hermaphrodites. These are comprised of exactly 959 cells, including 95 body wall muscle cells and 302 neurons that fall into 118 classes (WormBook 2016, Altun and Hall 2016). Its nervous system has been almost completely mapped by electron microscopy (White *et al* 1986). And it was the first multicellular animal whose genome had been completely sequenced (*C. elegans* Sequencing Consortium 1998). The nematode’s behavior and its underlying operation principles are the subject of numerous past and ongoing studies (Bono and Villu Maricq 2005, Corsi *et al* 2015), which have led to an extensive body of knowledge on this creature. This inspired biologists and neurocomputational researchers at the end of the last century to simulate not only the *C. elegans* nervous system, but the organism and its development in its entirety. We will point out some of the key works in this field to then focus on the scope and status of two concerted simulation initiatives, the OpenWorm and the *Si elegans* projects.

***C. elegans* as a model organism in neural computation**

With the advent of sufficiently powerful computational resources in the 80s of the last century, researchers discovered computers for the simulation of all kinds of natural phenomena, among them the events in nervous systems. Modeling neural systems has diverse roots and inspirations, most of them being inductively derived from first principles (e.g., (Hodgkin and Huxley 1952)) or deduced from direct observation. The nematode *C. elegans* was considered as an ideal system to start with.

Besides the pharyngeal system (Bhatla *et al* 2015), the most accessible circuit in *C. elegans* is its body wall muscle control system responsible for locomotion (Gjorgjieva *et al* 2014) consisting of 75 motor neurons (out of a total number of 113 motor neurons) of 8 classes that innervate 79 (out of 95) body wall muscle cells arranged posterior to the head along the dorsal and ventral cords (Riddle *et al* 1997, Gjorgjieva *et al* 2014, Zhen and Samuel 2015, Altun and Hall 2016). Although the nature of the driving neural signals is still under debate (electrotonic or graded-regenerative potentials *versus* self-terminating action potentials (Lockery and Goodman 2009, Lockery *et al* 2009)), its output can be

visualized rather easily and is thus verifiable and quantifiable by direct comparison with time-lapse recordings of worm postures (Stephens *et al* 2008, Brown *et al* 2013) and movements (Yemini *et al* 2013). Recent advances in designing genetically encoded optogenetic switches (Xu and Kim 2011) and fast-response calcium indicators (Gottschalk 2014) paired with real-time tracking (Faumont *et al* 2011), imaging techniques (Moy *et al* 2015) and laser ablation experiments (Gray *et al* 2005, Rakowski *et al* 2013) allow the association of postural changes with activity patterns and signal flow (Nguyen *et al* 2016, Venkatachalam *et al* 2016). Therefore, the majority of publications on simulating *C. elegans* focuses on various aspects of the sensory-motor loop (Lockery 2011, Cohen and Sanders 2014, Gjorgjieva *et al* 2014, Zhen and Samuel 2015) and its driving inputs (Schafer 2015).

Most of these studies were inspired by seminal work of Niebur and Erdős, who—based on reported behavioral descriptions of the nematode’s locomotion—laid the groundwork for the mathematical formalization of *C. elegans* nervous system function by taking body physics, environmental and biomechanical properties and resulting interaction forces into account (Niebur and Erdős 1991, Niebur and Erdős 1993). In 1992, a book entitled ‘AY’s Neuroanatomy of *C. elegans* for Computation’ was published that provided a neural circuitry database of the nematode *C. elegans* having been compiled from the literature that was available to that date (Achacoso and Yamamoto 1992). Several mathematical tools in form of BASIC and FORTRAN programs were made available that allowed the visualization and manipulation of morphological and circuit data to reveal correlations and functional roles. Despite the wealth of supplied data and the inspiring examples, it was cited more often as a general reference for small network architectures rather than serving as a basis for actual *C. elegans* network analyses studies. Since then, diverse strategies for describing neural events that lead to postural change have been proposed of which only a few are mentioned. Among them are event-driven models consisting of an asynchronous system based on pulse modulation (Claverol *et al* 1999), compartmental conductance-based models exclusively for muscle cells (Boyle and Cohen 2008), a central pattern generator that drives the forward movement of a physics-based rigid body representation of the nematode (Mailler *et al* 2010) inspired by (Niebur and Erdős 1991), neuromuscular control systems that rely on a sensory feedback mechanism based on bistable dynamics without the need for a modulatory mechanism except for a proprioceptive response to the physical environment (Boyle *et al* 2012, Williamson 2012), dynamic neural networks based on a differential evolution algorithm in the head and body with a central pattern generator in between acting on a locomotion model with 12 multi-joint rigid links (Deng and Xu 2014), evolutionary algorithms for the identification of a minimal klinotaxis network (Izquierdo and Beer 2013) and genetic algorithms to train 3680 synaptic weights within the motor connectome to replicate behaviors based on sensory-motor sequences (Portegys 2015) as recently reviewed by (Gjorgjieva *et al* 2014) and (Izquierdo and Beer 2016). The

lack of sufficient electrophysiological and biochemical data continues to fuel the connectome debate on whether the emergence of a certain behavior can be predicted solely from a network analysis (Jabr 2012, Seung 2012) featuring simplistic bistable neurons (Roberts *et al* 2016) or requires a more detailed description of neural events including other signaling modalities such as neuromodulators (Trojanowski and Raizen 2015) and proprioceptive (Butler *et al* 2015) or mechanosensory feedback (Bryden and Cohen 2004, Karbowski *et al* 2008).

Past named projects on simulating *C. elegans*

In 1997, researchers at the University of Oregon in the USA proposed ‘NemaSys’. It aimed at developing a computer simulation environment for *C. elegans* to support basic research and education in *C. elegans* and systems computational neuroscience. Due to *C. elegans*’s simplicity, an anatomically detailed model of the entire body and nervous system was perceived as an attainable goal. Over the years, a concerted effort employing electrophysiology, calcium imaging, quantitative behavioral analysis, laser ablation and mathematical modeling led to the identification of the mechanism and simple computational rules by which *C. elegans* computes the time derivative of chemosensory input (Ferree and Lockery 1999). The results were transcoded into a phototaxis response algorithm to control and analyze the trajectories of a custom-made robot (Morse *et al* 1998).

In 1998, ‘The Perfect *C. elegans* Project’, a collaboration between researchers with Sony, the Keio University in Japan and the University of Maryland in the USA, targeted at introducing synthetic models of *C. elegans* to further enhance our understanding of the underlying principles of its development and behavior, and life in general. Initial efforts focused on a realistic simulation of a subset of biological observables by providing a Java-based visualization tool for embryogenesis including cell position, kinematic interactions between cells, cell division, cell fate, neural connections and thermotaxis. Ultimately, a complete synthetic model of the nematode’s cellular structure and function, including genetic interactions, was envisioned. The concepts and first steps were outlined in an initial report (Kitano *et al* 1998), but were not followed up on.

In 2004, researchers at the Hiroshima and Osaka universities in Japan aimed at developing a virtual *C. elegans* in the ‘Virtual *C. elegans* Project’. Based on data on the spatial and structural layout of the nematode, they proposed a dynamic body model with muscles to analyze motor control. It was founded on a neural oscillator circuit to generate rhythmic movement. It could be shown that the model qualitatively generates rhythmic movements similar to wildtype and mutant nematodes. Another demonstration was a real-coded genetic algorithm to drive a kinematic locomotion model that responded to gentle-touch stimuli (Suzuki *et al* 2005a, 2005b).

Recent concerted project initiatives on simulating *C. elegans*

Emerging from the ‘CyberElegans’ project, the ‘OpenWorm’ project (USA, 2011-present; www.openworm.org) is an international open science project to simulate *C. elegans* from the cellular level upwards on standard and graphical processing unit (GPU)-enhanced computers via OpenCL. The long-term goal is to provide a full simulation of the *C. elegans* hermaphrodite. The first target is a description of the worm’s locomotion by simulating the 302 neurons and 95 body wall muscle cells (Szigeti *et al* 2014). Among the currently available modules is a realistic flexible worm body model including the muscular system and a partially implemented ventral neural cord (Palyanov *et al* 2011, Openworm Browser 2014). It is based on a merged and extended connectome dataset (Varshney *et al* 2011), which is editable in neuroConstruct (Gleeson *et al* 2007) and based on simulator-independent NeuroML standards (Gleeson *et al* 2010, Busbice *et al* 2012). Its visualization relies on the location dataset compiled by the ‘Virtual Worm Project’, an initiative at the California Institute of Technology that created an interactive atlas of the hermaphrodite’s cell-by-cell anatomy (Grove and Sternberg 2011). Locomotion can be described by a smoothed particle hydrodynamics simulator called ‘Siberetic’ (Palyanov *et al* 2016), which is similar to a system of spherical particles of different sizes that was reported to model both the nematode and its environment during movement and feeding behavior (Rönkkö and Wong 2008). Currently available open-source resources include the OpenWorm browser, the NeuroML *C. elegans* connectome, Siberetic and ‘Geppetto’ (Geppetto Contributors 2016), a web-based multi-algorithm, multi-scale simulation platform for simulating complex biological systems and their surrounding environment (Openworm Community 2016).

Around the same time, NEMALOAD (‘nematode upload’; USA, 2012–2014; github.com/nemaload) initiated the integration of a number of recent experimental imaging technologies (Marblestone *et al* 2013, Schrödel *et al* 2013) to learn how one neuron affects another in *C. elegans*. The project was structured in four subsequent stages that were supposed to build on one another. In the molecular biology stage, *C. elegans* strains should be functionalized with optogenetically encoded sensors and actuators (e.g., calcium indicators, photo-stimulators and inhibitors) for the tracing and manipulation of neural activity. In the imaging stage, this activity flow should be recorded in freely behaving worms at neuronal resolution. In the perturbation stage, individual neurons should be excited optically by means of a custom-made two-photon digital holography system to map their contributions to a certain behavior. In the final modeling stage, automation tools for the correlation of neural activity with behavior should have allowed the development of a dynamic model of the worm’s behavior in a simulated environment to mirror the experimentally observed behavior in its natural or laboratory environment. This should have allowed for elucidating the underlying information processing

structure. In 2014, these activities merged with the Open-Worm project.

The most recent concerted effort in emulating *C. elegans* is the *Si elegans* project (EU, 2013-present; www.si-elegans.eu). It aims at providing a closed-loop, open-source, peer-contribution platform being based on brain-mimetic principles for the emulation and reverse-engineering of *C. elegans* nervous system function in a behavioral context. *Si elegans* was motivated by the lack of a holistic closed-loop simulation environment, where neural events can be linked to as well as altered by their behavioral outcome. In this, the overall objectives are very similar to previous and ongoing endeavors. The chosen approach is slightly different, though. The nervous system consists of a dedicated hardware infrastructure that, unlike software implementations, permits true parallelism in the intra-neural as well as inter-neural signal processing. It is based on 329 field-programmable gate arrays (FPGAs), a parallel circuit definition architecture by design. Unlike functionally pre-defined neuromorphic computing systems (Furber 2016) based on very large-scale integrated circuit technology (Mead and Conway 1980), which—surprisingly—has not yet been exploited in the context of emulating *C. elegans* nervous system function, FPGAs are freely reconfigurable circuit fabrics that can accommodate distinct neural response models, just one for each *C. elegans* neuron (requiring 302 FPGAs) or several at a time. Similarly, FPGAs can carry one or several other models that interact with neurons, such as models of downstream muscle response (e.g., 27 FPGAs sharing up to 6 muscle models each to emulate the 95 *C. elegans* striated body wall muscles and 60 nonstriated muscles) and algorithms of subsequent body physics. These circuit-embedded response models may be dynamic and context-aware (Machado *et al* 2014, 2015, 2016) and thus evolve over time. This adaptation is not restricted to simply adjusting e.g., synaptic weights, but may allow the model to respond differently as a function of the (sensory) signal type and origin, environmental conditions (e.g., T) and their history, or of the local level of ‘neuromodulatory biochemical background’ at a given time. In view of the high number (359 200) of adaptive logic modules of the chosen FPGAs (Altera Stratix V GX), models are thus allowed to include aspects that are often ignored in computational neuroscience. For instance, the complexity of the dendritic tree suggests its involvement in the computational pre-processing of incoming signals such as their temporal filtering and amplitude modulation and its effect on altering synaptic properties (Smith 2010). Likewise, electrical junctions account for about 9% (recent work suggests up to 48% (Hall 2016)) of the overall interconnectivity between neurons, thereby constituting alternative signal propagation and—due to their permittivity for small molecules—modulation pathways to synaptic transmission.

Originally, the nematode’s entire connectome was to be implemented by a static light-projection scheme to warrant interference-free, parallel synaptic information transfer with high temporal fidelity (Petrushin *et al* 2014, 2015, Ferrara *et al* 2016). The axonal output of each FPGA neuron would have triggered a light-emitting diode, whose light was

projected through a patterned mask onto ‘synaptic’ photo-detectors of only those postsynaptic neurons that the respective presynaptic neuron connected to. This required each of the 302 light sources to carry a different neuron-specific projection mask. The final system features a dynamic version of such opto-electrical connectome based on digital light processing technology. The reconfigurable digital micro-mirror devices substitute the static projection masks to allow for exploring the impact of changes in neural inter-connectivity on neural information processing. Due to its complexity and costs, it is currently restricted to the synaptic signal transmission between the 20 neurons of the pharyngeal sub-network¹ as a proof-of-concept implementation. The remaining 279 neurons² exchange synaptic and gap-junction information through an Ethernet backbone. To nevertheless warrant the temporal parallelism inherent to biological networks and events, the hardware-based network will operate on a central clock (50 MHz). Some neural operations will require more FPGA ‘hardware clock’ cycles than others. At the cost of real-time operation, the supervising FPGA-based controller will thus ensure that all model operations of all neurons including the inter-neural signal transmission within a ‘biological clock’ cycle are completed before a new one starts. Any delays related to different lengths at the axonal arbor or synaptic properties can be incorporated in the respective neural models on the individual FPGAs.

Inspired by previous work on a closed-loop simulation framework for body, muscles and neurons (Voegtlin 2011), this biomimetic hardware nervous system emulation is controlling a virtually embodied and physically realistic representation of the nematode (via soft-body physics) in an equally realistic three-dimensional virtual behavioral arena (e.g., an agar Petri dish) (Mujika *et al* 2014, 2016). In there, the virtual *C. elegans* will encounter commonly tested stimuli (e.g., touch, chemicals, electric fields, light and/or temperature gradients) at any pre-defined time. These, together with characteristics of the environment (e.g., the shape of the plate, substrate properties) and the initial position and orientation of the nematode, can be batch-defined in a dedicated behavioral experiment configuration interface. The definition parameters are translated into an editable extensible markup language schema. During an experiment, the sensory experience is transmitted to the sensory neurons in the FPGA network. Based on published knowledge on network-internal circuitry and signal processing pathways, the sensory input (and proprioceptive information) will generate a motor output to instruct the muscles of the virtual worm on what to do next. In this closed-loop scenario, it will furthermore be possible to read out any network state (e.g., synaptic weights) at any given time for the reverse-engineering of network function. The simulation results, both the neuron variable traces as well as the body motion, can be visualized and downloaded after the simulation is over. To make the *Si elegans* framework

¹ The pharyngeal network is thought to be connected to the main network by only a single gap junction between I1 and RIP.

² CANL/R were excluded since they have no obvious synapses. VC6 was omitted as well since it only makes one neuromuscular junction.

user-friendly for novice and expert users alike, several model generation (e.g., drag-and-drop) and import functionality (e.g., from existing simulation engines) are provided (Krewer *et al* 2014, Morgan *et al* 2015). The current model design is based on the low entropy modeling specification language. In a neural network configuration graphical user interface, the user places neuron and synapse models in a graphically represented *C. elegans* connectome and can parametrize specific neuron models.

Once the chosen models generate a behavioral output that is comparable to observations in real laboratory experiments, the platform will allow the neuroscience community to better understand, if not anticipate, the neural mechanisms that underlie behavior. The open-source, peer-contribution *Si elegans* platform is publically accessible through platform.si-elegans.eu. Its early implementation and functionality may be compared with personal computers (PCs) in the 70s of the last century: just like the PC hardware and its basic operating system at that time, the *Si elegans* platform provides a basic computational framework to model *C. elegans* nervous system function and observe the generated behavioral output. Its usefulness in predicting neural function to reproduce a certain behavior will therefore strongly depend on its adoption and on contributions by both the biological and neurocomputational communities.

Discussion and conclusion

Biological nervous systems are robust and highly adaptive information processing entities that excel current von-Neumann-type computer architectures in almost all aspects of sensory-motor integration. While they are slow and inefficient in the serial processing of stimuli or data chains, they outperform artificial computational systems in seemingly ordinary pattern recognition, orientation or navigation tasks due to their parallel and multifactorial information processing capabilities. In terms of number of neurons and interneural connections, *C. elegans* is the most prominent and astonishing example of how a most minimalistic nervous system can process a multitude of different stimuli and sustain a diverse repertoire of behavioral outcomes. This irreconcilability suggests that there must be other mechanisms involved, which render this nervous system computationally more powerful beyond plugging a number of stereotypic computational units together. Yet, despite its seeming simplicity, *C. elegans* is keeping the third generation of biologists, neural engineers and computational neuroscientists busy in elucidating the underlying principles of how genes translate into nervous system function and a certain behavioral phenotype.

When Sydney Brenner proposed *C. elegans* as a model organism to the Medical Research Council in the UK in 1963, he stated that ‘We intend to identify every cell in the worm and trace lineages’ (Brenner 1963). While this goal has been achieved, it became clear that this information is insufficient to deduce the cells’ contributions to behavior. Several key questions are still unanswered. One of them is our lack of biological knowledge that would instruct us to what level of

detail a simulation has to drill down to let realistic behavior emerge. Will we need to uncover and formalize the entirety of the molecular machineries that underpin worm biology or will a more abstracted, thermodynamics-inspired description faithfully elicit the observed behavior *in silico*? Although we know most of the neurons’ roles and purposes (e.g., sensory, inter, motor, projection, local/solitary), still little is known about their identity (excitatory or inhibitory) and the relevance of the individual connections (including gap junctions). Furthermore, evidence suggests the existence of parallel, sometimes opposing (inhibitory versus excitatory) circuits. Similarly challenging are divergent circuits from a common starting point to different endpoints. In addition, the neural dynamics of different neurons are not uniform and even vary between individuals. Moreover, they may be modulated by extrasynaptic neural activation mechanisms including diffusible biochemical regulators (e.g., neuromodulators) or physical parameters (e.g., temperature, proprioception) (Bargmann and Marder 2013). These, in turn, may vary with internal states (e.g., starved versus satiated) and the environmental conditions. On top of that, synapses are constantly remodeled not only in response to behavioral experience, but in a context-sensitive and time- or activity-dependent manner on the timescale of milliseconds to weeks (Friston 2011). Thus, *C. elegans*’s neural circuit, despite its quasi-static wiring diagram, features many dynamic and difficult to capture mechanisms that encode different behavioral outcomes.

Due to this complexity and the many unknowns, any simulation approach is almost doomed to start with naïve and oversimplified assumptions. No matter how a simulation framework is conceptualized, the above findings strongly suggest to keep it as flexible, extensible and scalable as possible to accommodate new insights into the mechanisms that govern nervous system function underlying a particular behavioral phenotype. This may include the deviation from standard reasoning: instead of building population- or neuron-specific response models (Marder and Taylor 2011), an even more fine-grained approach may become necessary that provides a variety of adaptive models for one and the same neuron each responding to context-specific events. For this reason, new computational architectures such GPUs and FPGAs are explored to lift the restraints from the required hardware resources. In doing so, the OpenWorm and *Si elegans* initiatives both aim at providing the required tools in support of answering the question of how the *C. elegans* nervous system encodes behavior.

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References

- Achacoso T and Yamamoto W 1992 *AY's Neuroanatomy of C. elegans for Computation* (Boca Raton, FL: CRS Press)
- Altun Z F, Herndon L A, Wolkow C A, Crocker C, Lints R and Hall D H 2016 *Wormatlas: Handbook of C. elegans anatomy* <http://www.wormatlas.org>
- Bargmann C I and Marder E 2013 From the connectome to brain function *Nat. Methods* **10** 483–90
- Bhatla N, Droste R, Sando Steven R, Huang A and Horvitz H R 2015 Distinct neural circuits control rhythm inhibition and spitting by the myogenic pharynx of *C. elegans* *Curr. Biol.* **25** 2075–89
- Bono M D and Villu Maricq A 2005 Neuronal substrates of complex behaviors in *C. elegans* *Annu. Rev. Neurosci.* **28** 451–501
- Boyle J H, Berri S and Cohen N 2012 Gait modulation in *C. elegans*: an integrated neuromechanical model *Front. Comput. Neurosci.* **6** 10
- Boyle J H and Cohen N 2008 *Caenorhabditis elegans* body wall muscles are simple actuators *BioSystems* **94** 170–81
- Brenner S 1963 Proposal to the medical research council, appendix I—differentiation in a Nematode worm <http://hobertlab.org/how-the-worm-got-started/>
- Brown A E X, Yemini E I, Grundy L J, Jucikas T and Schafer W R 2013 A dictionary of behavioral motifs reveals clusters of genes affecting *Caenorhabditis elegans* locomotion *Proc. Natl Acad. Sci.* **110** 791–6
- Bryden J and Cohen N 2004 A simulation model of the locomotion controllers for the nematode *Caenorhabditis elegans* *From Animals to Animats 8: Proc. 8th Int. Conf. on the Simulation of Adaptive Behavior (Los Angeles, CA, 13–17 July 2004)* (USA: MIT Press) pp 183–92
- Busbice T, Gleeson P, Khayrulin S, Cantarelli M, Dibert A, Idili G, Palyanov A and Larson S 2012 The NeuroML *C. elegans* connectome *5th INCF Congress of Neuroinformatics (12 September 2012, Munich)*; *Front. Neuroinformatics* (doi:10.3389/conf.fninf.2013.08.00017)
- Butler V J, Branicky R, Yemini E, Liewald J F, Gottschalk A, Kerr R A, Chklovskii D B and Schafer W R 2015 A consistent muscle activation strategy underlies crawling and swimming in *Caenorhabditis elegans* *J. R. Soc. Interface* **12** 12
- C. Elegans* Sequencing Consortium 1998 Genome sequence of the nematode *C. elegans*: a platform for investigating biology *Science* **282** 2012–8
- Claverol E, Cannon R, Chad J and Brown A 1999 Event based neuron models for biological simulation. A model of the locomotion circuitry of the nematode *C. elegans* *Computational Intelligence and Applications* ed N E Mastorakis (Singapore: World Scientific) 217–22
- Cohen N and Sanders T 2014 Nematode locomotion: dissecting the neuronal–environmental loop *Curr. Opin. Neurobiol.* **25** 99–106
- Corsi A K, Wightman B and Chalfie M 2015 A transparent window into biology: a primer on *Caenorhabditis elegans* *Genetics* **200** 387–407
- Deng X and Xu J-X 2014 A 3D undulatory locomotion model inspired by *C. elegans* through DNN approach *Neurocomputing* **131** 248–64
- Desalle R and Tattersall I 2012 Do plants have brains? some biologists argue that ‘neurobiology’ has been too narrowly defined *Nat. Hist.* **120** (Online, available: <http://naturalhistorymag.com/features/152208/do-plants-have-brains>)
- Epstein H F and Shakes D C 1995 *Caenorhabditis Elegans: Modern Biological Analysis of an organism* (San Diego: Academic)
- Faumont S et al 2011 An Image-free opto-mechanical system for creating virtual environments and imaging neuronal activity in freely moving *Caenorhabditis elegans* *PLoS One* **6** e24666
- Ferrara L, Petrushin A, Liberale C, Brannick D, Connolly B, Mitchell P and Blau A 2016 Comparison of electro-optical strategies for mimicking *C. elegans* network interconnectivity in hardware *Biosystems & Biorobotics—Advances in Neurotechnology, Electronics and Informatics* ed A R Londral and P Encarnação (Cham: Springer)
- Ferree T C and Lockery S R 1999 Computational rules for chemotaxis in the nematode *C. elegans* *J. Comput. Neurosci.* **6** 263–77
- Friston K J 2011 Functional and effective connectivity: a review *Brain Connectivity* **1** 13–36
- Furber S 2016 Large-scale neuromorphic computing systems *J. Neural Eng.* **13** 051001
- Geppetto Contributors 2016 Geppetto: a web based platform to explore, visualize and simulate complex biological models <http://geppetto.org/>
- Gjorgjieva J, Biron D and Haspel G 2014 Neurobiology of *Caenorhabditis elegans* locomotion: where do we stand? *BioScience* **64** 476–86
- Gleeson P, Steuber V and Silver R A 2007 Neuroconstruct: a tool for modeling networks of neurons in 3D space *Neuron* **54** 219–35
- Gleeson P et al 2010 Neuroml: a language for describing data driven models of neurons and networks with a high degree of biological detail *PLoS Comput. Biol.* **6** e1000815
- Gottschalk A 2014 Optogenetic analyses of neuronal network function and synaptic transmission in *Caenorhabditis elegans* *e-Neuroforum* **5** 77–85
- Gray J M, Hill J J and Bargmann C I 2005 A circuit for navigation in *Caenorhabditis elegans* *Proc. Natl Acad. Sci. USA* **102** 3184–91
- Grove C A and Sternberg P W 2011 The virtual worm: a three-dimensional model of the anatomy of *caenorhabditis elegans* at cellular resolution *18th Int. C. elegans Meeting*
- Hall D H 2016 Gap junctions in *C. elegans*: their roles in behavior and development *Deve. Neurobiol.* advanced online publication (doi:10.1002/dneu.22408)
- Hodgkin A L and Huxley A F 1952 A quantitative description of membrane current and its application to conduction and excitation in nerve *J. Physiol.* **117** 500–44
- Izquierdo E J and Beer R D 2013 Connecting a connectome to behavior: an ensemble of neuroanatomical models of *C. elegans* *Klinotaxis PLoS Comput. Biol.* **9** e1002890
- Izquierdo E J and Beer R D 2016 The whole worm: brain-body-environment models of *C. elegans* *Curr. Opin. Neurobiol.* **40** 23–30
- Jabr F 2012 The connectome debate: is mapping the mind of a worm worth it *Sci. Am.* **5**
- Karbowski J, Schindelman G, Cronin C J, Seah A and Sternberg P W 2008 Systems level circuit model of *C. elegans* undulatory locomotion: mathematical modeling and molecular genetics *J. Comput. Neurosci.* **24** 253–76
- Kitano H, Hamahashi S and Luke S 1998 The perfect *C. elegans* project: an initial report *Artif. Life* **4** 141–56
- Krewer F, Coffey A, Callaly F and Morgan F 2014 Neuron models in FPGA hardware: a route from high level descriptions to hardware implementations *2nd Int. Congress on Neurotechnology, Electronics and Informatics (Rome, Italy)* (SCITEPRESS Digital Library) pp 177–83
- Lockery S R 2011 The computational worm: spatial orientation and its neuronal basis in *C. elegans* *Curr. Opin. Neurobiol.* **21** 782–90
- Lockery S R and Goodman M B 2009 The quest for action potentials in *C. elegans* neurons hits a plateau *Nat. Neurosci.* **12** 377–8
- Lockery S R, Goodman M B and Faumont S 2009 First report of action potentials in a *C. elegans* neuron is premature *Nat. Neurosci.* **12** 365–6
- Machado P, Wade J and McGinnity T M 2014 *Si elegans*: FPGA hardware emulation of *C. elegans* nematode nervous system

- 6th World Congress on Nature and Biologically Inspired Computing (NaBIC) 30 July 2014–1 Aug 2014) pp 65–71
- Machado P, Wade J and McGinnity T M 2016 *Si elegans*: Modeling the *C. elegans* nematode nervous system using high performance FPGAS *Biosystems & Biorobotics—Advances in Neurotechnology, Electronics and Informatics* ed R A Londral and P Encarnaçao (Cham: Springer International Publishing)
- Machado P, Wade J J, Appiah K and McGinnity T M 2015 *Si elegans: Hardware Architecture and Communications Protocol Int. Joint Conf. on Neural Networks (IJCNN)* (Killarney, Ireland) pp 3473–9
- Mailler R, Avery J, Graves J and Willy N 2010 *A Biologically Accurate 3D Model of the Locomotion of Caenorhabditis Elegans 2010 Int. Conf. on Biosciences (BIOSCIENCESWORLD)* pp 84–90
- Marblestone A H et al 2013 Physical principles for scalable neural recording *Front. Comput. Neurosci.* **7**
- Marder E and Taylor A L 2011 Multiple models to capture the variability in biological neurons and networks *Nat. Neurosci.* **14** 133–8
- Mead C and Conway L 1980 *Introduction to VLSI Systems* (Reading, MA: Addison-Wesley)
- Morgan F, Krewer F, Callaly F, Coffey A and Ginley B M 2015 Web-enabled neuron model hardware implementation and testing *3rd Int. Congress on Neurotechnology, Electronics and Informatics (Lisbon, Portugal, 16–17 November 2015)* ed A Faisal et al (Portugal: SCITEPRESS Digital Library) pp 138–45
- Morse T M, Lockery S R and Ferrée T C 1998 Robust spatial navigation in a robot inspired by chemotaxis in *Caenorhabditis elegans* *Adapt. Behav.* **6** 393–410
- Moy K, Li W Y, Tran H P, Simonis V, Story E, Brandon C, Furst J, Raicu D and Kim H 2015 Computational methods for tracking, quantitative assessment, and visualization of C-elegans locomotory behavior *PLoS One* **10** 22
- Mujika A, Epelde G, Leškovský P and Oyarzun D 2016 Physically-based simulation and web visualization of *C. elegans* behavioural experiments *Biosystems & Biorobotics—Advances in Neurotechnology, Electronics and Informatics* ed R A Londral and P Encarnaçao (Cham: Springer)
- Mujika A, Epelde G, Mauro A D and Oyarzun D 2014 Visualization of a virtual *Caenorhabditis elegans* in WebGL *2nd Int. Congress on Neurotechnology, Electronics and Informatics (Rome, Italy, 25 October 2014 through 26 October 2014)* (SCITEPRESS Digital Library) pp 164–8
- Nguyen J P, Shipley F B, Linder A N, Plummer G S, Liu M, Setru S U, Shaevitz J W and Leifer A M 2016 Whole-brain calcium imaging with cellular resolution in freely behaving *Caenorhabditis elegans* *Proc. Natl Acad. Sci.* **113** E1074–81
- Niebur E and Erdos P 1993 Theory of the locomotion of nematodes: control of the somatic motor neurons by interneurons *Math. Biosci.* **118** 51–82
- Niebur E and Erdös P 1991 Theory of the locomotion of nematodes: dynamics of undulatory progression on a surface *Biophys. J.* **60** 1132–46
- Openworm Browser 2014 <http://browser.openworm.org/>
- Openworm Community 2016 OpenWorm for Scientists <http://openworm.org/science.html>
- Palyanov A, Khayrulin S and Larson S D 2016 Application of smoothed particle hydrodynamics to modeling mechanisms of biological tissue *Adv. Eng. Softw.* **98** 1–11
- Palyanov A, Khayrulin S, Larson S D and Dibert A 2011 Towards a virtual *C. elegans*: a framework for simulation and visualization of the neuromuscular system in a 3D physical environment *In Silico Biol.* **11** 137–47
- Petrushin A, Ferrara L and Blau A 2015 The *Si elegans* connectome: a neuromimetic emulation of neural signal transfer with DMD-structured light *SPIE Photonics West, Emerging Digital Micromirror Device Based Systems and Applications VII, Session 6: Novel and Emerging DMD Applications* ed M R Douglass et al (San Francisco: SPIE)
- Petrushin A, Ferrara L, Liberale C and Blau A 2014 Towards an electro-optical emulation of the *C. elegans* connectome *2nd Int. Congress on Neurotechnology, Electronics and Informatics (Rome, Italy)* (SCITEPRESS Digital Library) pp 184–8
- Portegys T E 2015 Training sensory–motor behavior in the connectome of an artificial *C. elegans* *Neurocomputing* **168** 128–34
- Rakowski F, Srinivasan J, Sternberg P W and Karbowski J 2013 Synaptic polarity of the interneuron circuit controlling *C. elegans* locomotion *Front. Comput. Neurosci.* **7** 128
- Riddle D L, Blumenthal T, Meyer B J and Priess J R 1997 *Introduction: The Neural Circuit for Locomotion* (New York: Cold Spring Harbor Laboratory Press)
- Roberts W M et al 2016 A stochastic neuronal model predicts random search behaviors at multiple spatial scales in *C. elegans* *eLife* **5** e12572
- Rönkkö M and Wong G 2008 Modeling the *C. elegans* nematode and its environment using a particle system *J. Theor. Biol.* **253** 316–22
- Schafer W R 2015 Mechanosensory molecules and circuits in *C. elegans* *Pflügers Archiv—Eur. J. Physiol.* **467** 39–48
- Schrödel T, Prevedel R, Aumayr K, Zimmer M and Vaziri A 2013 Brain-wide 3D imaging of neuronal activity in *Caenorhabditis elegans* with sculpted light *Nat. Methods* **10** 1013–20
- Seung S 2012 *Connectome: How the Brain's Wiring Makes Us Who We Are* (Mariner Books)
- Smith L S 2010 Neuromorphic systems: past, present and future *Brain Inspired Cognitive Systems 2008* ed A Hussain et al (New York: Springer)
- Stephens G J, Johnson-Kerner B, Bialek W and Ryu W S 2008 Dimensionality and Dynamics in the Behavior of *C. elegans* *PLoS Comput. Biol.* **4** e1000028
- Suzuki M, Goto T, Tsuji T and Ohtake H 2005a A dynamic body model of the nematode *C. elegans* with neural oscillators *J. Robot. Mechatronics* **17** 318–26
- Suzuki M, Tsuji T and Ohtake H 2005b A model of motor control of the nematode *C. elegans* with neuronal circuits *Artif. Intell. Med.* **35** 75–86
- Szigeti B, Gleeson P, Vella M, Khayrulin S, Palyanov A, Hokanson J, Currie M, Cantarelli M, Idili G and Larson S 2014 OpenWorm: an open-science approach to modelling *Caenorhabditis elegans* *Front. Comput. Neurosci.* **8** 15
- Trojanowski N F and Raizen D M 2015 Neural circuits: from structure to function and back *Curr. Biol.* **25** R711–3
- Varshney L R, Chen B L, Paniagua E, Hall D H and Chklovskii D B 2011 Structural properties of the *Caenorhabditis elegans* neuronal network *PLoS Comput. Biol.* **7** e1001066
- Venkatachalam V et al 2016 Pan-neuronal imaging in roaming *Caenorhabditis elegans* *Proc. Natl Acad. Sci. USA* **113** E1082–8
- Voegtlin T 2011 CLONES: a closed-loop simulation framework for body, muscles and neurons *BMC Neurosci.* **12** 363
- White J G, Southgate E, Thomson J N and Brenner S 1986 The structure of the nervous system of the nematode *Caenorhabditis elegans* *Phil. Trans. R. Soc. B* **314** 1–340
- Williamson D R 2012 Modelling the locomotion nervous system in the nematode *C. elegans*: a developmental perspective *MSc Thesis* University of Leeds
- WormBook 2016 A comprehensive, open-access collection of original, peer-reviewed chapters covering topics related to the biology of *Caenorhabditis elegans* and other nematodes <http://wormbook.org/>
- Xu X and Kim S K 2011 The early bird catches the worm: new technologies for the *Caenorhabditis elegans* toolkit *Nat. Rev. Genet.* **12** 793–801

Yemini E, Jucikas T, Grundy L J, Brown A E X and Schafer W R 2013 A database of *Caenorhabditis elegans* behavioral phenotypes *Nat. Methods* **10** 877–9

Zhen M and Samuel A D T 2015 *C. elegans* locomotion: small circuits, complex functions *Curr. Opin. Neurobiol.* **33** 117–26