



Allergic rhinitis

Jean Bousquet^{1,2,3,4}✉, Josep M. Anto^{5,6,7,8}, Claus Bachert^{9,10,11,12}, Ilaria Baiardini¹³,
Sinthia Bosnic-Anticevich^{14,15}, G. Walter Canonica¹³, Erik Melén¹⁶, Oscar Palomares¹⁷,
Glenis K. Scadding¹⁸, Alkis Togias¹⁹ and Sanna Toppila-Salmi²⁰

Abstract | Allergic rhinitis (AR) is caused by immunoglobulin E (IgE)-mediated reactions to inhaled allergens and is one of the most common chronic conditions globally. AR often co-occurs with asthma and conjunctivitis and is a global health problem causing major burden and disability worldwide. Risk factors include inhalant and occupational allergens, as well as genetic factors. AR impairs quality of life, affects social life, school and work, and is associated with substantial economic costs. The Allergic Rhinitis and its Impact on Asthma (ARIA) initiative classified AR into intermittent or persistent and mild or moderate/severe. The diagnosis is based on the clinical history and, if needed in patients with uncontrolled rhinitis despite medications or with long-lasting symptoms, on skin tests or the presence of serum-specific IgE antibodies to allergens. The most frequently used pharmacological treatments include oral, intranasal or ocular H₁-antihistamines, intranasal corticosteroids or a fixed combination of intranasal H₁-antihistamines and corticosteroids. Allergen immunotherapy prescribed by a specialist using high-quality extracts in stratified patients is effective in patients with persistent symptoms. Real-world data obtained by mobile technology offer new insights into AR phenotypes and management. The outlook for AR includes a better understanding of novel multimorbidity phenotypes, health technology assessment and patient-centred shared decision-making.

Allergic rhinitis (AR) is characterized by sneezing, nasal congestion, nasal itching and rhinorrhoea (nasal discharge) and is caused by immunoglobulin E (IgE)-mediated reactions to inhaled allergens. These immune reactions involve mucosal inflammation that is driven by type 2 cells^{1,2}. AR seems to be the consequence of environmental exposures acting on a predisposed genetic background. AR is often co-morbid with asthma and/or conjunctivitis.

AR is one of the most common chronic conditions in high-income countries, with a prevalence of up to 50% in some countries³. By contrast, the prevalence is relatively low in low-income and middle-income countries (LMICs), although prevalence is increasing steadily in these countries. AR is a global health problem that causes major burden and disability worldwide. Indeed, AR contributes to missed or unproductive time at work⁴ and school, sleep problems and, in children, decreased involvement in outdoor activities⁵. The economic effect of AR is often underestimated as indirect costs are substantial, but the effect of AR on work productivity is estimated to cost €30 billion to €50 billion per year in the European Union^{4,6,7}.

The diagnosis of AR is made by medical history and examination (physical examination and, if needed, nasal endoscopy) plus, in some patients, tests for

allergen-specific IgE (skin prick tests or tests for serum-specific IgE). Available treatments include allergen avoidance, pharmacotherapy with H₁-antihistamines or intranasal corticosteroids (INCS) and allergen-specific immunotherapy (AIT). Many patients are dissatisfied with their treatment, for example, because management does not take the patient's needs into consideration, no cure is available, adherence to long-term therapy is poor and/or because the patient does not fully understand the condition. Real-world data obtained via mobile technology should offer new insights into the phenotypes and management of AR.

This Primer discusses the epidemiology, risk factors and genetic background of AR. In addition, it reviews the mechanisms of disease, diagnosis, prevention and management and the effect of AR on quality of life (QOL).

Epidemiology Prevalence

For accurate prevalence estimates, AR needs to be distinguished from infectious rhinitis and non-allergic rhinitis. Two large multinational studies on the prevalence of allergic diseases were conducted from 1990 to 2010 in children (International Study of Asthma and Allergy in Childhood (ISAAC))⁸ and adults (European Community

✉e-mail: jean.bousquet@orange.fr
<https://doi.org/10.1038/s41572-020-00227-0>

Author addresses

¹Charité – Universitätsmedizin Berlin, Humboldt-Universität zu Berlin, Berlin, Germany.

²Berlin Institute of Health, Comprehensive Allergy Center, Department of Dermatology and Allergy, Berlin, Germany.

³University Hospital Montpellier, Montpellier, France.

⁴MACVIA-France, Montpellier, France.

⁵ISGlobAL, Centre for Research in Environmental Epidemiology, Barcelona, Spain.

⁶IMIM (Hospital del Mar Research Institute), Barcelona, Spain.

⁷Universitat Pompeu Fabra, Barcelona, Spain.

⁸CIBER Epidemiología y Salud Pública, Barcelona, Spain.

⁹Upper Airways Research Laboratory, ENT Department, Ghent University Hospital, Ghent, Belgium.

¹⁰Sun Yat-sen University, International Airway Research Center, First Affiliated Hospital Guangzhou, Guangzhou, China.

¹¹Division of ENT Diseases, CLINTEC, Karolinska Institutet, Stockholm, Sweden.

¹²Department of ENT Diseases, Karolinska University Hospital, Stockholm, Sweden.

¹³Personalized Medicine Clinic Asthma & Allergy, Humanitas University & Research Hospital-IRCCS, Rozzano, Milan, Italy.

¹⁴Woolcock Institute of Medical Research, University of Sydney, Glebe, NSW, Australia.

¹⁵Woolcock Emphysema Centre and Sydney Local Health District, Sydney, NSW, Australia.

¹⁶Institute of Environmental Medicine, Karolinska Institutet and Sachs' Children's Hospital, Stockholm, Sweden.

¹⁷Department of Biochemistry and Molecular Biology, School of Chemistry, Complutense University of Madrid, Madrid, Spain.

¹⁸The Royal National ENT Hospital, University College Hospitals, London, UK.

¹⁹Division of Allergy, Immunology, and Transplantation (DAIT), National Institute of Allergy and Infectious Diseases, NIH, Bethesda, MD, USA.

²⁰Skin and Allergy Hospital, Karolinska Institutet, Helsinki University Hospital and University of Helsinki, Helsinki, Finland.

Health Survey (ECRHS)³; however, more recent large studies have not been undertaken. Collectively, these studies demonstrated that AR often begins early in life, with a prevalence of more than 5% at 3 years of age. In the ISAAC phase III study (consisting of data from 236 centres in 98 countries), AR prevalence increased from 8.5% in individuals aged 6–7 years to 14.6% in those aged 13–14 years⁸. Overall, AR in children and adolescents was more prevalent in high-income countries, but the prevalence of severe symptoms was higher in LMICs⁹. In the ECRHS study (conducted in 35 centres in 15 countries), the age-standardized and sex-standardized prevalence of AR in people aged 20–44 ranged from 11.8% in Oviedo (Spain) to 46.0% in Melbourne (Australia)³. Of note, AR is more common in males before puberty, whereas it is more common in females after puberty; these differences are more pronounced in those with asthma and AR concomitantly¹⁰. In the US National Health and Nutrition Examination Survey III from 1988 to 1994, the prevalence of nasal symptoms during the previous 12 months was highest (~30%) in individuals aged 17–29 years and lowest (~10%) in those older than 60 years¹¹.

In general, the prevalence of AR has increased worldwide since the 1960s in parallel to the increase in the prevalence of atopy (that is, the tendency to produce IgE antibodies owing to genetic and/or environmental factors)^{12,13}. The ISAAC also evaluated the change in prevalence of allergic diseases between 1994–1995 and 2002–2003 (REF.⁸). For the two age groups evaluated as part of this study (6–7-year-olds and 13–14-year-olds), the prevalence of AR increased from the 1990s to the early years of the first decade of the twenty-first century in many LMICs but decreased or had little change in

western Europe. The reasons for the observed changes in prevalence are unclear⁸.

Risk factors

Allergens associated with AR include pollens (tree, grass and weed, including ragweed), moulds and indoor allergens (house dust mites and animal allergens) and have a large geographical variability within and between countries¹⁴. Occupational AR includes both IgE (vegetal and animal proteins as well as certain chemicals) and non-IgE (isocyanates, persulfate salts and woods) mechanisms¹⁵. Risk factors for AR include antibiotic use, self-reported air pollution, exposure to farm animals (only in LMICs), exposure to cats and/or dogs, maternal and paternal smoking and vigorous physical activity in adolescents¹⁶. Of note, many of these risk factors are shared with asthma and atopic dermatitis¹⁶. Overweight and obesity are not associated with AR¹⁷. Of note, many of these exposures and lifestyle risk factors have not been established as major risk factors for AR¹⁸; for example, ambient air pollution and passive smoking do not seem to have a large effect on AR development, but pollution may be associated with increased AR severity¹⁹.

The proportion of rhinitis in general that is attributable to atopy is ~50% in the overall population²⁰. AR, asthma and atopic dermatitis often coexist in the same individual, partially due to a shared genetic origin^{21,22}. Indeed, data from genome-wide association studies (GWAS) have demonstrated that allergic diseases and traits share a large number of genetic susceptibility loci, of which *IL33*, *IL1RL1* (also known as *IL33R*), *IL13-RAD50*, *C11orf30* (also known as *EMSY*)–*LRR32* and *TSLP* seem to be important for multimorbid allergic diseases^{18,23}. In addition, rhinitis was associated with TLR expression, whereas AR associated with asthma was associated with *IL5* and *IL33*, suggesting a different genetic cause for AR alone compared with multimorbid AR²⁴. Further research is warranted to explore transcriptomic signatures as biomarkers for single and multimorbid allergic diseases.

Susceptibility loci for AR have various immune functions, such as the inflammatory adhesion process for *MRPL4* (19q13), in the activation, development and maturation of B cells and epithelial barrier function/regulatory T cell function for *BCAP* (also known as *PIK3API*; 10q24) and immune tolerance for *C11orf30-LRR32* (11q13), whereas other loci have unknown functions, such as *FERD3L* (7p21)²³. In addition, data from one large GWAS and HLA fine-mapping study found 20 new loci that were associated with AR, many of which had immune functions related to both innate and adaptive IgE-related mechanisms²⁵. In that study, the estimated proportion of AR attributable to the key identified AR-associated loci was 39%, which is a relatively high estimate for a complex disease. Other GWAS analyses have found common genetic mechanisms in AR and non-allergic rhinitis^{22,25,26}.

Classification

Although historically AR has been categorized as seasonal and perennial, this distinction has not been well reproduced in epidemiological studies assessing

molecular allergens as most patients are polysensitized (sensitized to more than one allergen)²⁷. Accordingly, the organization Allergic Rhinitis and its Impact on Asthma (ARIA) proposed replacing seasonality with intermittent and persistent rhinitis¹.

The large number of risk factors associated with AR suggest that the geographical variations in prevalence are due to a constellation of environmental factors varying between locations and time. These varying constellations of risk factors are also a plausible explanation for the time and place distribution of allergic multimorbidity¹⁶. No biomarker that can be used in clinical practice to predict the type (that is, phenotype or endotype) and severity of AR and the development of its common co-morbidities is available.

Multimorbidities and sensitization

Most patients with asthma have multimorbid rhinitis (AR or non-allergic rhinitis), whereas less than one-third of patients with AR have asthma associated with rhinitis¹. The Mechanisms of the Development of ALLergy (MeDALL) study, which included 12 European birth cohorts, demonstrated that the coexistence of rhinitis with asthma and/or atopic dermatitis is more common than expected by chance alone, both in the presence and in the absence of IgE sensitization, suggesting that multimorbidity and IgE have different genetic mechanisms²⁸. In addition, data from the MeDALL study suggested that type 2 signalling pathways represent a relevant multimorbidity mechanism of allergic diseases²².

Many patients with AR also have conjunctivitis, but rhinitis and rhinoconjunctivitis seem to be two separate diseases^{29–31} and should be differentiated. In addition, data from the mobile application MASK-air have identified an extreme pattern of uncontrolled multimorbidity (uncontrolled rhinitis, conjunctivitis and asthma during the same day)³². More recent classical epidemiological studies showed that ocular symptoms are more common in polysensitized patients²⁹ whether they have asthma or do not have asthma³⁰, ocular symptoms are associated with the severity of nasal symptoms³³, ocular symptoms are important to consider in severe asthma³³ and the severity of allergic diseases increases with the number of allergic multimorbidities³⁴.

Monosensitization and polysensitization represent different phenotypes of IgE-associated disease³⁵. Polysensitization is associated with an earlier onset of allergy and with more severe symptoms compared with monosensitization. In addition, the multimorbidity polysensitization phenotype seems to occur at various ages and in various allergenic environments and may be associated with specific mechanisms of disease³⁶.

Mechanisms/pathophysiology

Allergen exposure, either topically intranasally or in an exposure chamber³⁷, can be used to study nasal allergic reactions, monitor symptoms and collect nasal secretions and serum for mediator measurements in response to the allergen³⁸ or nasal scrapings or biopsy from patients^{39–41}. In addition, blood cells (such as basophils and antigen-specific T cells) and nasal mucosa

tissue can be studied *ex vivo*⁴² using different stimuli and interventions.

The nasal epithelium

The nasal mucosa is the primary air conditioner of the respiratory tract and the first line of defence against airborne infectious agents. For these roles, maintaining and restoring epithelial integrity and the ability to initiate immune responses are essential. In the presence of conditions or factors that impair mucosal integrity, the epithelium releases alarmins and other damage-associated molecular patterns that initiate repair mechanisms but can also induce protective inflammation. In AR, the same mechanisms may be active in inducing disease. For example, allergens with protease activity (such as Der p 1 in house dust mites) can directly compromise the epithelial barrier, whereas others (such as Der p 2 in house dust mites) can activate pattern recognition receptors; in both cases the epithelium can initiate innate immune responses through the release of alarmins such as IL-33, thymic stromal lymphopoietin (TSLP) or IL-25 (REFS^{43–46}). Alarmins can, in turn, activate group 2 innate lymphoid cells (ILC2s), which rapidly produce *in situ* type 2 cytokines (IL-5, IL-13 and IL-4), therefore having a key role in the initiation and maintenance of type 2 adaptive immune responses, leading to IgE class switching and mucosal inflammation. Additional environmental factors are probably involved in the pathophysiology of AR through their effects on the nasal epithelium. These include pollutants (diesel exhaust particles⁴⁷ or other air pollutants⁴⁸), irritants and infectious agents (*Staphylococcus aureus*⁴⁹ or viruses). The exact mechanisms through which those factors contribute to disease manifestation have not been elucidated.

Antigen presentation and sensitization

The allergic immune response begins with a sensitization phase when the patient is first exposed to an allergen without experiencing clinical symptoms (FIG. 1). During this phase, dendritic cells in the nasal mucosa take up the allergen, process it and transport it to the draining lymph node, in which the processed allergen is presented to naive CD4⁺ T cells. Following antigen presentation, naive CD4⁺ T cells are activated and differentiate into allergen-specific type 2 T helper cells (T_H2 cells)^{50–55} that induce the activation of B cells and IgE class switching, which leads to B cell differentiation into plasma cells that produce allergen-specific IgE. IgE enters the circulation and binds through its Cε3 domain to the high-affinity IgE receptor (FcεRI) on the surface of effector cells (for example, mast cells and basophils). These processes lead to the formation of a pool of memory allergen-specific T_H2 cells and B cells. Although this constitutes the first step in the development of allergy, it is also an ongoing process as the mucosa becomes exposed to various allergens on a chronic, seasonal or episodic basis.

Symptom generation and inflammation

AR symptoms are caused by biochemical products released in the nasal tissue during an allergic reaction. When a patient who has been sensitized by previous exposure to the allergen re-encounters the causative

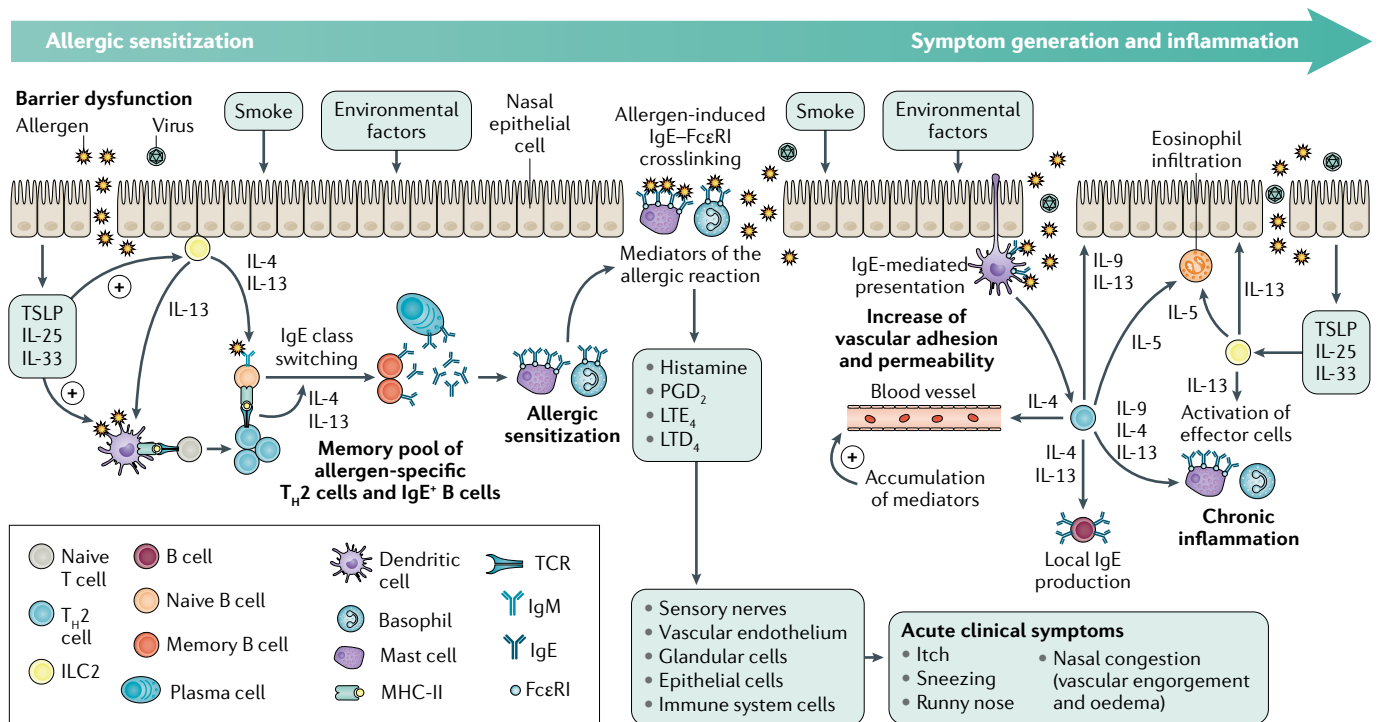


Fig. 1 | **Pathophysiology of allergic rhinitis.** During the sensitization phase, allergens are taken up by dendritic cells in the nasal mucosa and induce a series of events leading to the generation of plasma cells that produce allergen-specific immunoglobulin E (IgE) that binds to mast cells and basophils and to a pool of memory allergen-specific type 2 T helper cells (T_H2 cells) and IgE⁺ B cells. In individuals who are sensitized to the allergen, subsequent allergen exposure activates basophils and mast cells in the nasal mucosa, triggering the release of allergic mediators (including histamine and sulfidopeptide leukotrienes), leading to the acute symptoms of allergic rhinitis. Subsequent cytokine production by memory allergen-specific T_H2 cells induces an inflammatory infiltrate (eosinophil recruitment) within a few hours, leading to more symptoms and changes in the functional aspects of the nasal mucosa that mimic chronic rhinitis. ILC2, group 2 innate lymphoid cell; LTD₄, leukotriene D₄; LTE₄, leukotriene E₄; MHC-II, class II major histocompatibility complex; PGD₂, prostaglandin D₂; TCR, T cell receptor; TSLP, thymic stromal lymphopoietin. Adapted with permission from REF.⁵¹, Wiley.

allergen, the allergen binds to allergen-specific IgE on mast cells in the nasal mucosa, resulting in IgE and FcεRI crosslinking and subsequent mast cell activation and degranulation. This leads to the release of prestored and newly synthesized mediators, including histamine, sulfidopeptide leukotrienes (leukotriene C₄ and leukotriene D₄), prostaglandin D₂ and other products⁵² (FIG. 1). These mediators interact with nasal sensory nerves, vasculature and glands, resulting in acute AR symptoms.

In addition to acute symptoms, experimental nasal exposure to an allergen in an individual with AR produces immediate signs of inflammation, such as plasma exudation and the development of a type 2 inflammatory infiltrate characterized by eosinophils, neutrophils and basophils and by a mononuclear infiltrate (primarily T_H2 cells)⁵⁶. Indeed, classic type 2 cytokines — such as IL-4, IL-5 and IL-13 — can be detected in tissue and measured in nasal secretions several hours after allergen exposure^{57,58} (FIG. 1). In the natural presentation of AR, the histopathology is quite similar to that after experimental allergen exposure, and activation of allergen-specific memory T_H2 cells by dendritic cells and other antigen-presenting cells such as B cells via mechanisms partially depending on IgE-facilitated allergen presentation is believed to play a critical role^{49,50}. Activated allergen-specific T_H2 cells produce large

amounts of IL-4, IL-5 and IL-13 that contribute to vascular permeability, the infiltration of eosinophils and other inflammatory cells into the nasal mucosa, local IgE production, increased mucus production, vascular leakage and expansion, and activation and differentiation of different subsets of T_H2 cells^{40,42,43}. Of particular interest are antigen-specific T_H2 peripheral blood cells that display markers such as CCR4, CD161 and CCR4 with very little expression of CD27. These cells, also known as T_H2α cells, are associated with AR severity and are characteristically suppressed by allergen immunotherapy⁵⁷. Although the histology of AR shows a typical type 2 inflammation, there is little evidence of mucosal remodelling in contradistinction with asthma^{46,52}.

Although patients with AR experience acute symptoms on exposure to a known allergen, the natural symptomatic state can be chronic with relatively low fluctuations and frequently includes non-allergen triggers such as irritants and changes in environmental conditions. Experimental allergen provocations (nasal challenges), whether conducted by direct instillation of an allergen into the nasal cavities or through an environmental exposure unit, help identify a number of pathophysiological phenomena that offer better understanding of the overall clinical picture of AR. Acute symptoms are reduced within minutes but can persist

at lower levels for hours after allergen exposure or can recrudescence in a phenomenon known as the ‘late-phase’ reaction⁵⁹. Late-phase reactions may explain why, after sudden exposure to a large amount of an allergen, a patient with AR may remain symptomatic for prolonged periods. Repetitive allergen exposure leads to another phenomenon, whereby progressively lower amounts of the allergen are required to elicit symptoms; this has been termed ‘priming’⁶⁰. Nasal priming may explain why, towards the end of a pollen season, patients with AR tend to become symptomatic even when exposed to very low levels of pollen. A third phenomenon is the induction of nasal hyper-responsiveness, where the nasal response (sneezing, rhinorrhoea and so on) to a stimulus that is not an allergen (for example, histamine) can be augmented by a prior allergen challenge. In the natural history of AR, this phenomenon may explain the high sensitivity of patients to irritants and to environmental changes⁶¹.

Nasal priming may be caused by the increased accumulation of mast cells or basophils — which is a consequence of repeated allergen exposure — at the site of the allergic reaction in the nose⁶², in addition to the induction of non-specific hyper-responsiveness at the ‘end-organ’ (nose) level. In addition, late-phase reactions and chronicity might also depend on the perpetuation of allergen-specific adaptive immune responses. Indeed, persistent symptoms following allergen challenge could be mediated mainly by the accumulation of allergic mediators in the nasal mucosa but also by the activation of allergen-specific memory T_H2 cells by dendritic cells and other antigen-presenting cells such as B cells via mechanisms partially depending on IgE-facilitated allergen presentation^{63,64}. Activated T_H2 cells produce large amounts of type 2 cytokines that contribute to enhance all the local pathophysiological mechanisms described above, including the enhanced and sustained activation not only of tissue-resident mast cells but also of basophils infiltrating the nasal mucosa, which also might well contribute to the clinical symptoms after allergen-induced IgE crosslinking and subsequent release of mediators^{50–52} (FIG. 1). Nasal late-phase reactions, priming and non-specific hyper-responsiveness are inflammation dependent and can be suppressed by nasal corticosteroids. However, the specific molecular and cellular events underlying these phenomena are not yet fully understood.

Role of nasal nerves

The nervous system has a key role in the nasal symptoms of AR. The nasal mucosa is densely innervated by adrenergic and cholinergic nerve fibres, and the epithelium is innervated by interdigitating sensory nerve endings, mostly nociceptors (receptors that respond to noxious stimuli)⁶⁵. In addition, nasal chemosensory cells that express bitter taste receptors may reflect a specialized sensory nerve system that reacts to noxious stimuli and bacterial products⁶⁶. Cholinergic and adrenergic nerve fibres are activated through central reflexes initiated at the nasal mucosa by sensory nerves, including nociceptor C fibres. Nasal C fibres and other sensory nerves express receptors for some mediators of the allergic

response (such as histamine and bradykinin), and also express several transient receptor potential ion channels that are activated by noxious physical or chemical stimuli, such as low pH, high or low temperatures, CO₂ and hypertonicity. These fibres may also have a local effector function as they produce and release neuropeptides via axonal, antidromic reflexes⁶⁷. However, the mechanism and clinical role are unclear. Nasal hyper-responsiveness seems to have a strong neural component⁶⁸. Indeed, changes in the density and neuropeptide content of sensory nerves have been found in the nose of patients with AR. A putative mediator of these changes, nerve growth factor, is released on nasal allergen provocation⁶⁹ and can be found in nasal glandular epithelial cells and eosinophils in the nasal mucosa⁷⁰.

Diagnosis, screening and prevention

AR is often under-recognized owing to poor public awareness, limited access to allergologists and confounding diagnoses, such as the common cold⁷¹. The diagnosis of AR is made by considering a detailed history that is supported by examination findings (physical examination and, if needed, nasal endoscopy) and, if necessary, testing for allergen-specific IgE. Other tests such as nasal allergen challenge, CT scans, evaluation of nasal nitric oxide and ciliary beat frequency, nasal smears, nasal cultures and analysis of nasal fluid for β-transferrin) may be required to include or exclude different forms of rhinitis⁷². The ARIA guidelines propose classifying AR as intermittent or persistent depending on the duration of symptoms, with persistent rhinitis occurring for more than 4 days for 4 weeks at a time, and as mild or moderate to severe, depending on whether sleep and daily activities are affected or whether symptoms are troublesome⁷³ (FIG. 1). In addition, AR can also be classified as mild, moderate or severe⁷⁴.

Clinical history

The clinical history should note symptoms, particularly those that cause major problems, where and when they occur, and any exacerbating and relieving factors. Other symptoms in the chest, ears, throat, gut or skin, in addition to whether there is a patient history or a family history of allergic disease and/or immune problems, together with a review of treatments previously tried, those currently being taken and their efficacy, should all be noted.

All individuals are familiar with rhinitis because of the common cold. Rhinitis is defined clinically as having two or more of the following symptoms for more than 1 hour per day: nasal running, blocking, itching or sneezing. The diagnosis is made by an accurate history but can be missed owing to misperceptions such as symptoms being ascribed to frequent colds, mouth-breathing children having enlarged adenoids and secretions being missed when they pass posteriorly. Questioning patients with asthma regarding nasal function (such as the ability to breathe through the nose and to smell) should be undertaken as most patients with asthma have rhinitis or rhinosinusitis⁷³.

The clinical history may also provide a clue to the inciting allergen or allergens. Of note, long-term allergen

exposure is harder to diagnose than short-term exposure as the major symptom is often nasal blockage and postnasal discharge, with fewer obvious symptoms such as nasal itching, running and sneezing. Nasal inflammation can also cause non-specific nasal hyper-reactivity to non-allergic stimuli⁷⁵ and a poor sense of smell, among other multimorbidities⁷⁶. Some pollen-sensitive individuals with AR may present with oral symptoms of pollen food syndrome, such as itchy mouth and throat after ingestion of the food⁷⁷.

Examination

An examination of the whole patient is necessary as rhinitis has important co-morbidities⁷⁶. Children should have their growth assessed, as severe airway problems are associated with reduced growth, and the combined use of INCS and inhaled corticosteroids may reduce height at high doses⁷⁸. The presence of facial features such as conjunctivitis, nasal allergic crease, allergic salute or double creases beneath the eyes (Dennie–Morgan lines) all suggest that the patient has an allergic diathesis.

Nasal examination is needed in patients with moderate to severe AR or in those with uncontrolled symptoms despite optimal treatment. This examination should include assessment of the external appearance followed by internal examination, preferably with a nasendoscope. An otoscope may suffice to examine the nose in children. The position of the nasal septum, as well as the size and colour of the inferior turbinates, should be noted, together with the appearance of the mucosa and the presence and nature of any secretions, polyps, bleeding, tumours, crusting or foreign bodies. The classic appearance of the nasal cavity in patients with AR is swollen pale bluish inferior turbinates with copious clear secretions; however, the nose may look normal, and these features are not restricted to AR. Of note, the mucosa may be slightly reddened in patients using INCS⁷². Referral to an ear, nose and throat specialist is advised for patients with nasal polyps, bleeding, unilateral disease, high crusting and septal perforations⁷⁹. High crusting and septal perforations most commonly result from previous septal surgery but can also occur with cocaine abuse and vasculitides.

Asthma should always be assessed in patients with AR by asking patients about wheezing, shortness of breath and sleep disturbance plus, if needed, an objective measurement such as spirometry⁸⁰ and vice versa in patients with asthma owing to the frequent co-occurrence of these disorders. Patients should also undergo ear inspection as otitis media with effusion (also known as glue ear) may be a co-morbidity in children with rhinitis and in adults with severe forms of rhinosinusitis⁸¹. The general examination needs to also include skin examination for atopic dermatitis, and, in patients with obstructive rhinitis, assessment of thyroid function by checking for slow relaxation after the ankle jerk and for eye signs such as puffiness, redness and/or bulging, as hypothyroidism.

Tests

Whether further diagnostic testing for AR is required in all patients is disputed. Some clinicians do not recommend further testing in those with a clear history of nasal

symptoms that are provoked by allergen exposure⁸². This occurs in most northern hemisphere patients allergic to pollen and with intermittent exposure to animal or occupational allergens but is not possible with long-term exposure, as with pollens in most tropical climates or indoor allergens such as house dust mites. By contrast, other clinicians recommend further testing in all patients with symptoms suggestive of AR. Testing for allergen-specific IgE using skin prick or blood tests⁷² to identify the allergen can be performed to support the diagnosis and is mandatory when AIT is being considered as part of the treatment. Of note, the results from IgE testing need to be interpreted in the light of the clinical history, as both false-positive and false-negative results can occur⁸³. In one meta-analysis of skin prick tests, the sensitivity ranged from 68% to 100% and the specificity ranged from 70% to 91% (REF.⁸⁴).

Component-resolved diagnosis (that is, using purified native or recombinant allergens to identify IgE sensitivity to individual allergens) is not yet routine in AR diagnosis but may provide important information. For example, the detection of serum IgE antibodies to specific molecules (Phl p 1, Phl p 5, Bet v 1 or Pru p 3) could be used as a biomarker to predict AR persistence and the future onset of multimorbidities, such as asthma and/or pollen food syndrome^{85–87}. Moreover, component-resolved diagnosis may be useful in understanding cross-sensitizations and in proposing AIT where specific molecular sensitization can guide the content of the vaccine⁸⁶.

Other diagnostic tests may be required to verify the diagnosis of AR or to make an alternative diagnosis. Additional tests include nasal allergen challenge, nasal cytology, nasal nitric oxide measurements and ciliary beat frequency analysis⁷². Nasal smear cytology is practised in some centres. The presence of a high number of eosinophils (although the necessary percentage of cells is debatable) suggests an inflammatory process but not necessarily AR (AR or non-allergic rhinitis with eosinophilia), which is likely to be corticosteroid responsive. Unilateral eosinophilia can occur so bilateral samples must be taken⁸⁸. Nasal nitric oxide measurement is a simple and rapid test to discriminate AR, non-allergic rhinitis, and subgroups with acceptable sensitivity and specificity, but it can be done only in highly specialized centres⁸⁹.

Testing for local AR

Some patients with rhinitis who do not have systemic IgE sensitization identified via a skin prick test and serum allergen-specific IgE show nasal reactivity on a nasal allergen provocation test (whereby an allergen is entered into the nose). Nasal allergen provocation tests can be used when the history and systemic IgE results are not concordant, to monitor the progress of AIT and in research studies⁹⁰. Local AR might be an independent rhinitis phenotype, although it is treated in the same way as AR⁹¹.

Patients with rhinitis who have negative results on tests for allergy have non-allergic rhinitis, which can be caused by several factors that can broadly be divided into infectious and non-infectious factors. Tests to identify these factors are detailed in an EAACI position paper⁷².

Box 1 | Managing patients with AR during the COVID-19 pandemic

The global spread of COVID-19 has caused sudden and dramatic changes in society and health care, including management of allergic rhinitis (AR). Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) particles that are inhaled through the nose or the mouth bind to upper respiratory tract cells, and the nose is the first organ to be invaded²⁰³. In addition, SARS-CoV-2 binds to angiotensin-converting enzyme 2 (ACE2), and expression of ACE2 is lower in patients with allergic diseases, suggesting that they may be less prone to SARS-CoV-2 infection^{204,205}. The effect of AR on COVID-19 is still a matter of debate, and no firm conclusion can be drawn²⁰⁶. Smell dysfunction and anosmia are common COVID-19 symptoms²⁰⁷.

The allergy and immunology communities have quickly responded by mobilizing practice adjustments and embracing new paradigms of care to protect patients and staff from severe SARS-CoV-2 exposure^{208–211} using telehealth²¹². Some recommendations regarding the treatment of AR in patients with COVID-19 have been made. For example, although oral corticosteroids are contraindicated in patients with COVID-19, they can be used without restriction in patients with AR and COVID-19 (REF.²¹³). Practical recommendations were made to organize an allergy clinic²⁰⁸. In addition, Allergic Rhinitis and its Impact on Asthma (ARIA) and EAACI recommend stopping all forms of allergen-specific immunotherapy in patients with AR who have COVID-19 and continuing allergen-specific immunotherapy in patients with AR who do not have COVID-19 (REFS^{214–216}). The possibility of expanding injection intervals in the continuation phase should be checked and may be beneficial. On the other hand, face masks may reduce the severity of AR symptoms²¹⁷.

As for any other chronic disease, it is clear that the COVID-19 pandemic will profoundly change AR management owing to the infectivity of allergic patients, probable changes in management and major changes in health-care systems.

Mobile health

Mobile health tools use algorithms created using advanced statistics (neural networks) on data including medical diagnoses and questionnaire answers⁹². Very large numbers of patients can be studied with data obtained by mobile phones, and, in the future, mobile health added to machine learning may be useful for the screening of undiagnosed patients with AR in the general population.

Prevention

Many different attempts have been made to prevent allergic diseases, although most of these attempts have been unsuccessful. However, farm animal exposure in early life is a protective factor for allergic diseases in high-income countries⁹³ and also possibly in some LMICs, although the mechanisms are unclear⁹⁴. In addition, early-life exposure to cats and dogs may prevent the development of allergy but results are not consistent⁹⁵. The use of probiotics or prebiotics prenatally and postnatally has failed to reduce AR^{96,97}. Moreover, the use of pharmacological therapies before allergen exposure cannot prevent the onset of symptoms in AR⁹⁸.

Management

Treatments for AR include education, allergen avoidance, pharmacotherapy and AIT^{1,79,99}. Pharmacotherapy is effective in most patients and, when properly implemented, improves QOL; however, many patients do not follow prescriptions and are poorly adherent to pharmacotherapy.

No biologic has been approved for AR except omalizumab in Japan for *Cryptomeria japonica* allergy¹⁰⁰, and, owing to their costs and the prevalence of AR, newly developed biologics may be restricted to highly stratified patients with severe AR. Of note, some changes

to the management of AR have occurred owing to the COVID-19 pandemic (BOX 1).

Education

As AR is a chronic condition that is caused by specific allergens, it is important for patients to try to identify allergens and/or environmental agents that may precipitate their AR. In addition, it is important for clinicians to inform patients of the importance of the correct use of intranasal sprays to ensure they correctly adhere to the therapy.

Allergen avoidance

When possible, avoiding or minimizing exposure to the causative allergens should be the first management step for AR. Although allergen avoidance may reduce symptoms of AR, it should never isolate individuals from social interactions. Data regarding the benefits of house dust mite avoidance in asthma are conflicting, and interventions designed to reduce house dust mites have unknown effectiveness¹⁰¹. For patients who are polysensitized, avoidance strategies are often challenging as it is difficult to eliminate all causative allergens or triggers¹⁸. Some classical, but often unproven allergen avoidance measures for AR include use of bed covers for house dust mite allergy, removing pets from the home, changing profession for occupational allergens or wearing masks for pollen allergy. An innovative approach is to feed cats with a cat food containing antibodies against Fel d 1 to reduce allergenicity of the cat¹⁰².

Pharmacotherapy

Many pharmacological treatment options are available for the management of AR (TABLE 1). Oral and/or intranasal H₁-antihistamines, INCS and the fixed combination of INCS and H₁-antihistamines are all considered first-line treatments depending on the severity or burden of symptoms.

H₁-antihistamines. H₁-antihistamines for AR treatment are available in oral, intranasal and ocular formulations and are first-line treatments for patients with mild symptoms or those who do not want to use an INCS treatment. H₁-antihistamines block the action of histamine by acting as neutral receptor antagonists or inverse agonists of the histamine H₁ receptor¹⁰³. Different H₁-antihistamines have different chemical structures, pharmacokinetics and potential for drug–drug and drug–food interactions, and they are classified into non-sedating, less-sedating and sedating groups on the basis of brain H₁ receptor occupancy¹⁰³. The less-sedating second-generation oral H₁-antihistamines (such as desloratadine, loratadine, cetirizine, levocetirizine and rupatadine) and the non-sedating H₁-antihistamines (such as fexofenadine and bilastine) are well tolerated, safe and effective¹⁰⁴. First-generation oral H₁-antihistamines should be avoided owing to adverse effects, in particular sedation, and are not recommended for AR treatment^{103,105}. First-generation H₁-antihistamines have not been optimally studied as most trials of these therapies do not meet appropriate standards in terms of study design, meaning their relative efficacy is unknown¹⁰⁶.

Table 1 | Treatment options for allergic rhinitis

Treatment	Rhinorrhoea	Sneezing	Nasal itch	Nasal obstruction	Ocular symptoms	Onset of action
Oral H ₁ -antihistamine	++	++	+	+	+	1–3 hours
Intranasal H ₁ -antihistamine	++	++	+	+	0	<30 minutes
Ocular H ₁ antihistamine	0	0	0	0	+++	15 minutes
Intranasal corticosteroid	+++	+++	+++	+++	+ to ++	6–48 hours
Intranasal corticosteroid plus intranasal H ₁ -antihistamine	++++	++++	++++	++++	+++	10–60 minutes
Nasal decongestant	0	0	0	+++	0	15 minutes
Intranasal chromone	+	+	+	+	0	15 minutes
Ocular chromone	0	0	0	0	++	15 minutes
Leukotriene receptor antagonist	+	+	+	+	0	1 hour
Intranasal anti-cholinergic agent	++	0	0	0	0	1 hour

0, no evidence of efficacy; + to +++++, increasing levels of evidence of efficacy.

In general, the advantages of oral H₁-antihistamines are once-a-day administration, rapid and effective action and low cost. However, they are less effective than INCS, particularly for nasal congestion (which is a common symptom of AR). Oral H₁-antihistamines are often sufficient for the treatment of mild AR, and many patients prefer oral drugs to other formulations. Some oral H₁-antihistamines may, with caution, be used in pregnancy or in women who are breastfeeding (for example, cetirizine, levocetirizine and loratadine¹⁰⁷).

Topically administered H₁-antihistamines (alcaftadine, azelastine, bepotastine, cetirizine, epinastine, ketotifen, and olopatadine (as eye drops) and azelastine and olopatadine (as nasal sprays)) are suitable for patients with mild AR or ocular symptoms. These medicines may have a bitter taste in 15–20% of patients. The effect begins 1–3 hours after administration^{104,108}. Mild uncommon adverse effects include dry mouth, drug-induced rash and swelling of the salivary glands. As previously mentioned, patients should be educated on the correct way to administer intranasal H₁-antihistamines.

Intranasal corticosteroids. INCS such as beclomethasone, budesonide, ciclesonide, fluticasone propionate, fluticasone furoate, mometasone furoate and triamcinolone acetonide are first-line therapeutic options for patients with persistent or moderate to severe symptoms. INCS effectively control the four major symptoms of AR (TABLE 1), and some INCS (such as intranasally administered fluticasone furoate) can reduce ocular symptoms¹⁰⁹. INCS are more effective than H₁-antihistamines and leukotriene receptor antagonists, particularly for nasal congestion, although their efficacy requires several hours or days^{104,110} (TABLE 1). The mechanism of action of INCS is related to the local anti-inflammatory effect on nasal mucosal cells. As with intranasal formulations of other drugs, all patients should be educated on the correct way to administer intranasal products.

INCS are not systemically absorbed; therefore, there are no systemic adverse effects. The most common INCS adverse effects are local, including nasal irritation,

stinging and epistaxis¹¹¹, and can usually be prevented by aiming the spray slightly away from the nasal septum. Long-term INCS use does not damage nasal mucosa or induce glaucoma¹¹², and growth effects in children seem to be minimal⁷⁸. Some INCS, such as budesonide, can be safely used during pregnancy at the recommended therapeutic dose after a thorough medical evaluation¹¹³.

INCS and intranasal H₁-antihistamine fixed combination. Fixed-dose combinations of INCS and intranasal H₁-antihistamine for treatment of AR and rhinoconjunctivitis include fluticasone propionate–azelastine¹¹⁴ and mometasone–olopatadine, which was approved in Australia in 2019 (REF.¹¹⁵). These medications are more effective than the individual compounds administered separately, are well tolerated (except for some bitter taste in a few patients) and are effective within minutes (fluticasone propionate–azelastine)¹¹⁶ or within 1 hour (mometasone–olopatadine)¹¹⁷ (TABLE 1). These therapies are typically used in patients who failed to benefit from INCS treatment alone and have been suggested for use in non-adherent patients who treat their symptoms intermittently. Combining oral H₁-antihistamines and INCS does not seem to increase the efficacy of INCS^{118,119}.

Other drugs. Leukotriene receptor antagonists, montelukast and zafirlukast, are used in the treatment of AR. Their effect is close to that of oral H₁-antihistamines¹⁰⁴. In Europe, leukotriene receptor antagonists have been approved by the EMA only for patients with co-morbid asthma and AR. Other AR therapies, such as chromones and ipratropium bromide, are effective for only some symptoms. For example, chromones (disodium cromoglycate) can mostly be self-administered and tried in patients with mild local ocular symptoms¹⁰⁴. Chromones are safe, but their effectiveness is usually quite modest. The ipratropium bromide nasal spray is well tolerated but is effective only for nasal secretion¹⁰⁴. Decongestants include intranasal sprays (for up to 7 days), such as oxymetazoline or phenylephrine sprays, and H₁-antihistamines combined with decongestant

sympathomimetic tablets or capsules (for up to 10 days), such as acrivastine, cetirizine hydrochloride or desloratadine plus pseudoephedrine¹⁰⁴, and are indicated only in those with severe nasal obstruction and should not be used long term to avoid rhinitis medicamentosa (for intranasal preparations). Saline irrigation may reduce patient-reported disease severity in adults and children with AR, with no reported adverse effects^{120,121}. Herbal products, homeopathy and acupuncture are still largely used for treatment of AR but lack clear evidence, and herbal medicine can cause adverse effects such as contact dermatitis, headache, itchy eyes and gastrointestinal symptoms¹²².

AR pharmacotherapy and children. Few studies have evaluated AR pharmacotherapy in preschool children⁹⁹. The therapies with demonstrated efficacy are rupatadine¹²³ and, in school-aged children, cetirizine¹²⁴, azelastine hydrochloride and the fluticasone propionate–azelastine fixed combination¹²⁵. Cetirizine is approved at 6 months of age in some countries. In addition, INCS can be prescribed in preschool children, H₁-antihistamines are suitable for persons older than 1 year, and cromoglycate or antihistamine eye drops are suitable for patients older than 3 years.

AR pharmacotherapy in elderly patients. In most elderly patients, rhinitis symptoms, diagnosis and treatment are the same as for other adult age groups. However, elderly patients often have mixed AR and non-allergic rhinitis with multiple triggers and have higher levels of mucosal dryness than younger patients¹²⁶. Rhinitis symptoms in elderly patients may include profuse rhinorrhoea without itching, isolated nasal obstruction usually when lying down, and nasal crusting in winter or in patients treated with diuretics.

INCS, oral H₁-antihistamines, intranasal H₁-antihistamines and the azelastine hydrochloride–fluticasone propionate fixed combination are first-line therapeutic options in elderly patients¹²⁷. Of note, ageing can affect the nasal mucosa by increasing cholinergic

activity and atrophy¹²⁶; thus, the dose of topically administered therapies may need to be reduced in elderly patients. In addition, some therapies can cause specific adverse effects in elderly individuals. Indeed, oral decongestants or systemic glucocorticosteroids are not recommended in elderly patients owing to adverse effects¹²⁷. Oral decongestants can cause, for example, palpitations, insomnia, nervousness, irritability, trouble with urination and reduced appetite, whereas glucocorticosteroids can induce glaucoma, cataract, osteoporosis and diabetes mellitus. Moreover, first-generation H₁-antihistamines are strongly discouraged in elderly individuals owing to sedation and anticholinergic effects^{128,129}. Caution should be exercised in patients with co-existing diseases, poly medication and organ (such as renal or liver) dysfunctions¹²⁷.

Real-life data and next-generation guidelines. Real-life observational studies using mobile health have found that most patients with AR self-medicate or use over-the-counter medications, placing the pharmacist at the forefront of treatment¹³⁰. Patients consulting primary care physicians usually have uncontrolled symptoms despite use of multiple medications. Adherence to treatment is a major issue¹³¹, as many patients do not seek advice from physicians, do not follow the physician's prescriptions and self-medicate to control their symptoms often using over-the-counter medications¹³⁰. Surprisingly, the use of multiple medications is associated with poor rhinitis control¹³².

Some recommendations for AR treatment are based on the Grading of Recommendations, Assessment, Development and Evaluation (GRADE) guidelines^{104,118,119,133}. Next-generation guidelines¹³⁴ were subsequently developed using existing GRADE-based guidelines and real-world evidence including data from randomized controlled trials, real-world data provided by mobile technology^{131,132} and data from additive studies, such as allergen chamber studies assessing the speed of onset of medications^{116,117} (TABLE 2). Real-life data

Table 2 | Next-generation ARIA guidelines

Guideline	GRADE evidence (RCT)	Real-world evidence
Oral H ₁ -antihistamines are less potent than INCS	V	RCT and RWD
Many patients prefer oral drugs	None	None
Intranasal H ₁ -antihistamines are less effective than INCS	V	RCT and RWD
Intranasal H ₁ -antihistamines are effective within minutes	None	Chamber studies
INCS are potent medications, first-line therapy in moderate to severe AR	V	RCT and RWD
INCS need a few hours to 1–2 days to be effective (except ciclesonide)	V	Chamber studies
The combination of INCS and oral H ₁ -antihistamines does not offer advantage over INCS	V	RCT and RWD
INCS and intranasal H ₁ -antihistamines are more potent than INCS in moderate to severe AR	With restriction	RCT and RWD
INCS and intranasal H ₁ -antihistamines are effective between 10 and 60 minutes	None	Chamber studies

Adapted with permission from REF.¹³⁴, Elsevier. AR, allergic rhinitis; ARIA, Allergic Rhinitis and its Impact on Asthma; GRADE, Grading of Recommendations, Assessment, Development and Evaluation; INCS, intranasal corticosteroids; RCT, randomized controlled trial; RWD, real-world data using mobile technology.

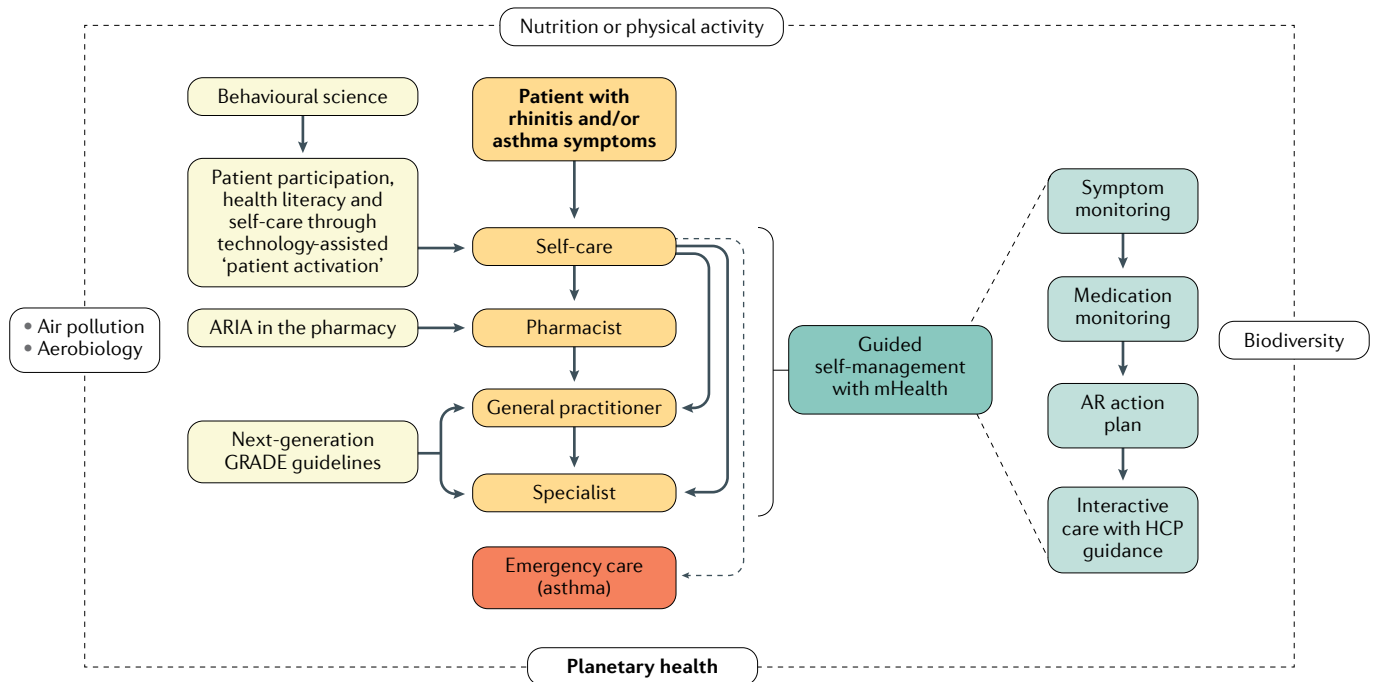


Fig. 2 | Care pathways for allergic rhinitis. The MASK-air study is conducted in 27 countries across the world. The MASK-air good practice was recognized by the Directorate-General for Health and Food Safety of the European Commission as a digital tool for citizen empowerment and for person-centred care. It proposes a stepwise care pathway for allergic rhinitis (AR). Patients with rhinitis symptoms typically use self-care. Patients typically then consult a pharmacist (in many but not all countries), then their general practitioner and then, if needed, specialists. Patients attend emergency care facilities in cases of severe asthma exacerbation. The next-generation care pathway approach for AR management proposes strategies to improve the stepwise approach using guided self-management with mobile health technology (mHealth). Non-pharmacological treatment (allergen and environmental pollutant avoidance, nasal saline douching exercise, nutrition and increasing biodiversity) needs to be included in the care pathway to sustain both patient and planetary health. ARIA, Allergic Rhinitis and its Impact on Asthma; GRADE, Grading of Recommendations Assessment, Development and Evaluation; HCP, health-care professional. Adapted from REF.¹³⁵, CC BY 4.0.

clearly indicate that patients prefer as-needed treatment to continuous treatment, and this should be reflected in future guidelines.

Care pathways for a digitally enabled, patient-centred approach. AR treatment should be individualized according to the symptom profile, severity and duration, the patient's preference of oral versus intranasal administration, and the availability and affordability of medications. Mobile technology can improve shared decision-making, and one example has been recognized as a Good Practice by the Directorate-General for Health and Food Safety of the European Commission for digital health in AR¹³⁵ and for change management¹³⁶ (FIG. 2).

Multistakeholder care pathways should be established in each country or region as health-care systems differ^{135,137}. As many patients with AR self-medicate and have poor symptomatic control, there is a great need to improve self-management for patients with AR, which can be aided by mobile health. In this scenario, patient counselling is first done by pharmacists with the help of allergy guides and websites (FIG. 3), following which patients should be referred to physicians. Mobile technology could reduce the time between the first symptoms and referral of patients with uncontrolled symptoms to specialists, as primary care physicians can

have an objective assessment after the first allergy season (pollen or indoor allergens), including adherence to treatment and control of rhinitis and asthma.

Allergen-specific immunotherapy

AIT is indicated for AR, allergic rhinoconjunctivitis and/or asthma when symptoms remain uncontrolled with avoidance measures and appropriate pharmacotherapy in adherent patients¹³⁸.

The aim of AIT is to induce tolerance to the allergens and, therefore, to reduce the symptoms of allergic diseases. For a sustained effect, AIT should be applied for a minimum of 3 years, either continuously or pre-seasonally¹³⁹. The induction of tolerance by AIT leads to changes in allergen-specific memory T cell and B cell responses and in the allergen-specific IgE and IgG antibody levels, and modifies the activation thresholds for mast cells, basophils and dendritic cells¹⁴⁰. Of note, the levels of allergen-specific nasal and serum IgG4 antibodies correlate closely with the clinical response to AIT in patients with AR¹⁴¹.

Selecting allergens that have a clinical effect on the patient is paramount to the success of AIT. These causative allergens can be identified through clinical history taking, component-resolved diagnosis and, if indicated and necessary, nasal provocation testing. The use of

prescription databases has indicated that the product-specific efficacy demonstrated in double-blind, placebo-controlled, randomized trials translates into real life¹⁴². Evidence of efficacy of AIT has been demonstrated for grass, birch (covering the homologous group of Betulaceae tree pollen), ragweed and *Cryptomeria japonica* pollen^{143,144}, as well as house dust mites¹⁴⁵, whereas less evidence is available for other types of pollen, animal dander or moulds. Only regulated, standardized allergen extracts that have demonstrated efficacy and safety should be used for AIT^{138,146,147}. However, efficacy can be assumed for allergens within homologous groups, including several pollen and house dust mite extracts as defined by respective EMA guidelines for allergen products¹³⁸. There is no evidence that mixing different allergens is effective in AIT, as this may result in underdosing and a potential degradation of specific allergens.

AIT can be applied via the subcutaneous or the sublingual routes, as tablets or drops, following the same indications and contraindications¹⁴⁸. Natural allergens or chemically modified allergens (known as allergoids) may be used, with the aim of reducing the risk of adverse events but maintaining efficacy or enabling an increased dosage^{138,149}. International and national guidelines are available^{104,150–152} and are updated on a regular basis.

Owing to the need for long-term use and the cost in most countries, only selected patients should receive AIT, which should be prescribed by allergists (FIG. 4). However, no validated biomarkers for predicting or monitoring the efficacy of AIT at an individual patient level are available in clinical practice¹⁵³, although mobile health may be of great interest. The symptom–medication score, grading symptoms and medication use on a daily basis, still remains the most reliable parameter of success in daily practice.

Adverse effects of AIT are relatively common but are rarely severe¹³⁸. Local reactions include redness and swelling at the injection site that occurs immediately or several hours after injection. Other adverse effects, such as sneezing, nasal congestion or hives, indicate systemic reactions. Serious reactions such as swelling of lips and tongue, laryngeal oedema, shortness of breath and chest tightness (asthma) in response to injections are very rare but require immediate medical attention and upfront preparations, such as availability of equipment and medications, and training of the personnel. Symptoms of an anaphylactic reaction typically include swelling in the throat, wheezing or tightness in the chest, nausea and dizziness and should be immediately treated with adrenaline (auto-injector) and preparation of an intravenous access. As most serious reactions develop within 30 minutes after injection, it is recommended that patients are supervised in the physician's office for at least 30 minutes before leaving. Allergen drops or tablets have a more favourable safety profile than injections. The first sublingual therapy dose should be administered under the supervision of a physician, but subsequent doses can be administered at home. Most adverse events are local (mouth itching, lip swelling and nausea) and spontaneously subside with further administration.

Quality of life

Health-related QOL (HRQOL) is the most frequently patient-reported outcome in AR (TABLE 3). The ARIA recommendations⁷³ have proposed grading AR severity by taking into account the effect of AR on HRQOL. Moreover, regulatory authorities such as the FDA¹⁵⁴ and the EMA¹⁵⁵ have provided guidance to the industry on how to use patient-reported outcomes to support labelling claims and routinely consider patient-reported outcomes as a tool used for data collection. In addition,

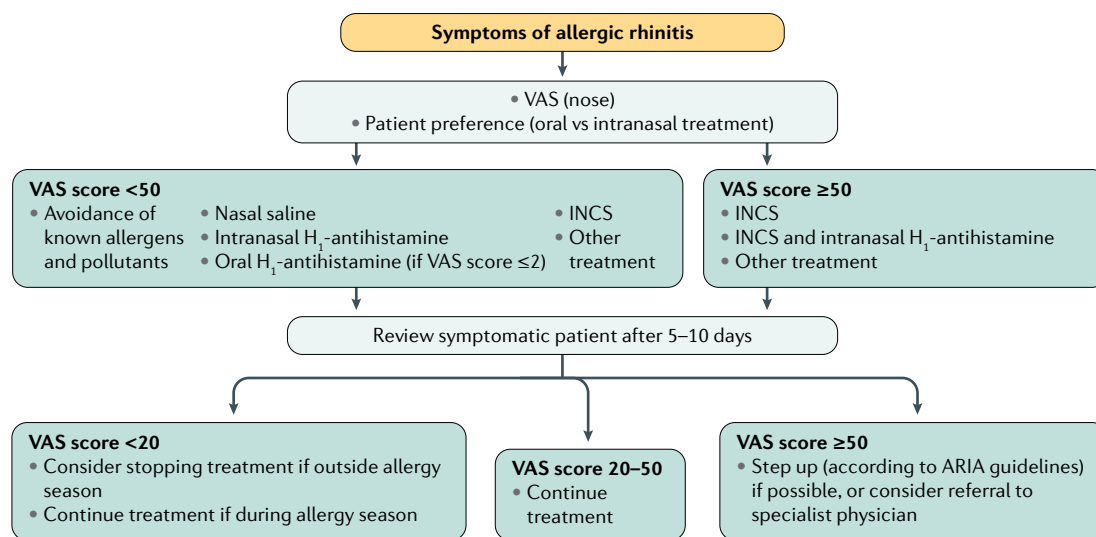


Fig. 3 | **Care pathway for the management of allergic rhinitis in the pharmacy.** A simple algorithm to help the pharmacist diagnose allergic rhinitis and dispense over-the-counter medications is required. Use of a visual analogue scale (VAS) to determine symptom severity is simple and accurate. The patient's symptoms need to be reassessed regularly if not daily to propose stopping the treatment, to continue use of over-the-counter medications or to send the patient to a physician. ARIA, Allergic Rhinitis and its Impact on Asthma; INCS, intranasal corticosteroids. Adapted with permission from REF.¹³⁰, Wiley.

patient preferences and values are introduced as cornerstones in GRADE, which is the best option for grading clinical evidence and developing recommendations for diagnostic and therapeutic interventions¹⁵⁶. Although patient-reported outcomes are not explicitly included in the definition of personalized medicine, they represent a real opportunity for involving patients in each step of disease management¹⁵⁷.

The availability of validated questionnaires for AR has permitted the evaluation of the effect of this disease in adults, children and adolescents. The use of generic tools, which are applicable to all health conditions, has underlined that adults with AR who have HRQOL scores significantly lower than those of the general population have HRQOL scores that are lower than those of patients with asthma¹⁵⁸. In addition, children and adolescents with AR had lower HRQOL scores than healthy peers¹⁵⁹. The effect of AR on the physical domain of QOL was comparable in teenagers with AR and in those with asthma¹⁶⁰.

The aspects of HRQOL that are relevant for patients with AR have been identified by disease-specific questionnaires^{161,162}. A rich literature shows how the presence and severity of symptoms negatively affect daily activities, performance, sleep, physical and emotional status and social functioning at all ages¹⁶³; the effect of AR and multimorbid asthma^{164–166}; the possibility to minimize or delete AR impact¹⁶⁴; and the effect of symptomatic treatments¹⁶⁷ or specific immunotherapy. Of note, AR impairs QOL to a greater extent than moderate asthma¹⁵⁸ and significantly impairs work productivity⁴.

Interest in the patients' perspective is continuously growing in AR research. Nonetheless, the routine use of

HRQOL in clinical practice, which is encouraged owing to the potential to optimize disease management^{168,169}, remains limited. In the next few years, the questionnaires for assessing and monitoring AR HRQOL in individual patients need to be validated¹⁷⁰ and introduced into routine care.

Outlook

Mobile health

Multimorbidity in allergic airway diseases is well known⁷⁶, but a mobile application (MASK-air) created to assess how multimorbidity affects symptoms and severity has provided further findings³². Indeed, data from this mobile application have revealed that AR and rhinoconjunctivitis do not seem to be the same disease and have identified a pattern of uncontrolled multimorbidity in some patients (uncontrolled rhinitis, conjunctivitis and asthma on the same day)³². Data from such mobile applications are generating hypotheses that need confirmation in epidemiological studies. In this regard, differences between AR alone and AR associated with conjunctivitis were previously known²⁹ but epidemiological studies using data from mobile applications demonstrated that ocular symptoms are more common in patients with polysensitization³⁰, are associated with nasal symptom severity³³ and are important to consider in severe asthma³³. Moreover, the severity of allergic diseases increases with the number of allergic multimorbidities³⁴. This is the first example of a discovery of novel allergic phenotypes using a mobile health application confirmed by classic epidemiological studies. Other mobile tools have been proposed¹⁷¹ but few have been tested¹⁷². There is an urgent need to replicate existing data and to optimize mobile health for AR management in the digital transformation of health and care¹⁷³. An interesting approach will be to propose alerts for pollen¹⁷⁴, pollution or asthma exacerbations¹⁷⁵.

Mechanisms and multimorbidity

As previously mentioned, allergic diseases are heterogeneous; some patients have AR alone, whereas others have AR and asthma (with or without other allergic manifestations), although few patients have asthma alone. In addition, there are probably common genes associated with asthma and AR and specific genes associated with AR alone. Combining big data analyses (such as from the MASK-air application¹³⁵), classical epidemiological studies, *in silico* analysis, transcriptomics using microarray data (as exemplified in MeDALL^{22,24,35}) or RNA sequencing¹⁷⁶ has led to the reclassification of the mechanisms of allergic diseases. For example, polysensitization and multimorbidity represent the extreme allergic phenotype, starting early in life, and are associated with *IL5* and *IL33* activation. Notably, several of these and other genes associated with allergic multimorbidity point towards type 2 inflammation¹⁷⁷ and eosinophil activation²⁴. By contrast, rhinitis alone is associated with Toll-like receptor pathways, which have a key role in the innate immune system. Assessing these mechanisms in more detail may allow better understanding of the mechanisms of allergy and provide novel insights for prevention and treatment. Although it is well established that

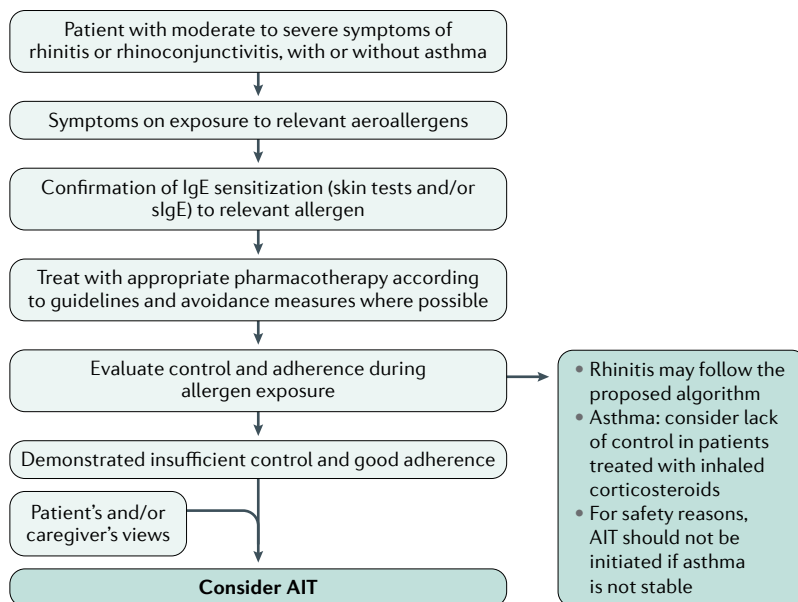


Fig. 4 | Stratification of patients for AIT. Only some patients with allergic rhinitis need allergen immunotherapy (AIT), such as patients who respond poorly to optimal treatment. Several steps are required before AIT can be performed, including a correct allergy diagnosis, the assessment of adherence to medications and the demonstration of an uncontrolled disease during the allergen exposure despite optimal medications. IgE, immunoglobulin E; sIgE, serum-specific immunoglobulin E. Adapted with permission from REF.¹³⁸, Wiley.

Table 3 | Questionnaires for the assessment of HRQOL in patients with rhinitis

Questionnaire	Number of items	Population	Ref.
Rhinoconjunctivitis Quality of Life Questionnaire (RQLQ)	28	Adults	196
raciborskiMini Rhinoconjunctivitis Quality of Life Questionnaire (Mini-RQLQ)	14	Adults	197
Nocturnal Rhinoconjunctivitis Quality of Life questionnaire (NRQLQ)	16	Adults	198
RHINASTHMA	30	Adults	199
RhinAsthma Patient Perspective (RAPP)	10	Adults	170
Adolescent Rhinoconjunctivitis Quality of Life Questionnaire	25	Adolescents	200
RHINASTHMA Adolescents	20	Adolescents	166
Pediatric Rhinoconjunctivitis Quality of Life Questionnaire (PRQLQ)	23	6–12 years	201
RHINASTHMA children	17	6–11 years	165
RhinAsthma Patient Perspective children (RAPP)	5	6–11 years	202

HRQOL, health-related quality of life.

rhinitis can lead to asthma, the exact phenotype of AR prone to developing asthma is still unclear. It is possible that polysensitized individuals can more commonly develop asthma.

New treatments

As many patients do not experience full relief from AR symptoms with available treatments, knowledge gaps undermine the development of new pharmacological and biological interventions to improve management. One important area of research is the identification of novel reliable biomarkers to phenotype or endotype patients with AR to predict treatment response and management strategies. Other areas of research include the elucidation of novel molecular mechanisms involved in allergen-specific response perpetuation in the nasal mucosa and translational integration of genomics, transcriptomics, proteomics and metabolomics into health care.

Novel cost-effective pharmacological treatments

Despite AR being one of the most prevalent diseases in the world with a high economic burden, no big pharmaceutical companies are developing novel treatments. The reasons for this paradox are complex but can be summarized as follows: overall, low-cost medications are effective in most patients if they are used appropriately, and, in many countries, they are given over the counter with no cost for the payers; AR is not a life-threatening disease and payers prioritize life-saving medications, for example, for cancer, rare diseases and COVID-19; developing medications with an efficacy substantially higher than that of those currently on the market may be difficult; and, owing to the number of patients with AR, only low-cost medications may eventually be reimbursed if patients with AR are not stratified, and therefore the cost of the development of an AR treatment largely surpasses potential revenues and very few trials are ongoing¹⁷⁸.

In addition, although monoclonal anti-IgE or anti-IL-4/IL-13 is effective in AR^{179,180} and some of the available biologics (or those in the pipeline) for asthma may be suitable for patients with severe, multimorbid allergic conditions or stratified patients with very severe

AR, there are only a few repurposing attempts for asthma medications. One example is montelukast, which, in Europe, is indicated only for asthma with AR multimorbidity as there was a request from the payers to lower the price of the asthma drug if the medication was also approved for AR alone. In addition, monoclonal antibodies to allergens may be of interest in those with AR caused by a single major allergen driving the allergic reaction.

Patient stratification is needed to find a group of patients that are unresponsive to the current medications and to obtain indirect costs incurred by these patients. The estimated cost of AR in Europe owing to presenteeism ranges from €25 billion to €50 billion⁶. A novel model of reimbursement of medications should be developed with, for example, enterprises paying for a potential new treatment with a precise cost-effectiveness analysis showing potential benefits. Mobile health can have a role in the cost-effectiveness analysis.

Improved treatment

Increasing safety while maintaining or even increasing efficacy are the main goals of research for novel vaccine development and improvement of treatment schemes in AIT. Perspectives in AIT are well established¹⁸¹, and many new products are in development^{182–185}. However, the vast majority of previous attempts failed because safety issues or lack of efficacy was observed. Future directions for conventional AIT include use of adjuvants, including vitamin D, Toll-like receptor ligand agonists, biologics¹⁸⁶ or probiotics¹⁸⁷. Several attempts have been made to increase tolerance and efficacy using molecular allergy vaccines acting on B cells or T cells but none has produced convincing results¹⁸⁸.

Treatment of AR could also be improved by the use of shared decision-making, a process whereby both the patient and the physician contribute to the medical decision-making process. In AR, data from mobile technology have revealed that patients are not adherent to treatment and self-medicate using as many medications possible to control their disease. Accordingly, there is an urgent need to propose shared decision-making using mobile health tools to optimize AR treatment. In addition, drug repurposing could be useful for treatment

of AR and could be aided by mobile health^{135,189,190}. Value-added medicine can help address unmet patient needs and could improve treatment-associated QOL.

The global allergy solution

The delivery of cost-effective modern health care is challenging for the management of chronic diseases and in particular for allergic diseases¹⁹¹, as management, which is often dependent on specialist and supporting services, is becoming unaffordable. Accordingly, innovative solutions often based on mobile health are required^{192,193}.

Authorities should be supported for the transformation of health and care towards integrated care with organizational health literacy for allergic diseases^{135,173,194}. A global allergy simple solution should provide the framework to digitally transform the prevention and control of allergic diseases in a cost-effective manner. Mobile health could be used to optimize an accessible and affordable treatment in stratified and participatory patients with allergic diseases to provide change management¹⁹⁵.

Published online: 03 December 2020

- Bousquet, J. et al. Allergic Rhinitis and its Impact on Asthma (ARIA) 2008 update. *Allergy* **63** (Suppl. 86), 8–160 (2008).
ARIA is a non-governmental organization that collaborated with the WHO. This authoritative review provides evidence that most patients with asthma have multimorbid rhinitis, whereas less than one-third of patients with AR have multimorbid asthma.
- Greiner, A. N., Hellings, P. W., Rotiroti, G. & Scadding, G. K. Allergic rhinitis. *Lancet* **378**, 2112–2122 (2011).
- Bousquet, P. J. et al. Geographical distribution of atopic rhinitis in the European community respiratory health survey I. *Allergy* **63**, 1301–1309 (2008).
- Vandenplas, O. et al. Impact of rhinitis on work productivity: a systematic review. *J. Allergy Clin. Immunol. Pract.* **6**, 1274–1286 (2018).
- Devillier, P. et al. In allergic rhinitis, work, classroom and activity impairments are weakly related to other outcome measures. *Clin. Exp. Allergy* **46**, 1456–1464 (2016).
- Zuberbier, T., Lotvall, J., Simoons, S., Subramanian, S. V. & Church, M. K. Economic burden of inadequate management of allergic diseases in the European Union: a GA²LEN review. *Allergy* **69**, 1275–1279 (2014).
- Colas, C. et al. Estimate of the total costs of allergic rhinitis in specialized care based on real-world data: the FERIN study. *Allergy* **72**, 959–966 (2017).
- Asher, M. I. et al. Worldwide time trends in the prevalence of symptoms of asthma, allergic rhinoconjunctivitis, and eczema in childhood: ISAAC phases one and three repeat multicountry cross-sectional surveys. *Lancet* **368**, 735–743 (2006).
ISAAC is the largest global prevalence study of allergic diseases. Its results have shown that AR prevalence increases from childhood to adolescence. From the 1990s to the early years of the first decade of the twenty-first century, prevalence was still increasing in many developing countries.
- Ait-Khaled, N. et al. Global map of the prevalence of symptoms of rhinoconjunctivitis in children: the international study of asthma and allergies in childhood (ISAAC) phase three. *Allergy* **64**, 123–148 (2009).
- Keller, T. et al. The sex-shift in single disease and multimorbid asthma and rhinitis during puberty - a study by MeDALL. *Allergy* **73**, 602–614 (2018).
- Singh, K., Axelrod, S. & Bielory, L. The epidemiology of ocular and nasal allergy in the United States, 1988–1994. *J. Allergy Clin. Immunol.* **126**, 778–783 (2010).
- Latvala, J., von Hertzen, L., Lindholm, H. & Haahntela, T. Trends in prevalence of asthma and allergy in Finnish young men: nationwide study, 1966–2003. *BMJ* **230**, 11–86–87 (2005).
- Eder, W., Ege, M. J. & von Mutius, E. The asthma epidemic. *N. Engl. J. Med.* **355**, 2226–2235 (2006).
- Wheatley, L. M. & Togias, A. Allergic rhinitis. *N. Engl. J. Med.* **372**, 456–463 (2015).
- Moscato, G. et al. Occupational rhinitis. *Allergy* **63**, 969–980 (2008).
- Asher, M. I. et al. Which population level environmental factors are associated with asthma, rhinoconjunctivitis and eczema? Review of the ecological analyses of ISAAC Phase One. *Respir. Res.* **11**, 8 (2010).
- Tajima, H. & Pawankar, R. Obesity and adiposity indicators in asthma and allergic rhinitis in children. *Curr. Opin. Allergy Clin. Immunol.* **19**, 7–11 (2019).
- Wise, S. K. et al. International consensus statement on allergy and rhinology: allergic rhinitis. *Int. Forum Allergy Rhinol.* **8**, 108–352 (2018).
In this critical review of the AR literature, the authors apply a systematic evidence-based approach to provide a comprehensive, clinical update on current knowledge of AR and to identify knowledge gaps for future studies.
- Burte, E. et al. Long-term air pollution exposure is associated with increased severity of rhinitis in 2 European cohorts. *J. Allergy Clin. Immunol.* **145**, 834–842 (2020).
- Zacharasiewicz, A., Douwes, J. & Pearce, N. What proportion of rhinitis symptoms is attributable to atopy? *J. Clin. Epidemiol.* **56**, 385–390 (2003).
- Ferreira, M. A. et al. Shared genetic origin of asthma, hay fever and eczema elucidates allergic disease biology. *Nat. Genet.* **49**, 1752–1757 (2017).
This landmark GWAS on allergic multimorbidity strongly suggest that asthma, hay fever and eczema partly coexist because they share many genetic risk variants that dysregulate the expression of immune-related genes.
- Anto, J. M. et al. Mechanisms of the Development of Allergy (MeDALL): introducing novel concepts in allergy phenotypes. *J. Allergy Clin. Immunol.* **139**, 388–399 (2017).
- Li, J., Zhang, Y. & Zhang, L. Discovering susceptibility genes for allergic rhinitis and allergy using a genome-wide association study strategy. *Curr. Opin. Allergy Clin. Immunol.* **15**, 33–40 (2015).
- Lemonnier, N. et al. A novel whole blood gene expression signature for asthma, dermatitis, and rhinitis multimorbidity in children and adolescents. *Allergy* <https://doi.org/10.1111/all.14314> (2020).
This MeDALL collaborative study represents the largest transcriptomics allergy dataset published to date and describes unique blood gene expression signatures in allergic multimorbidity versus rhinitis only.
- Waage, J. et al. Genome-wide association and HLA fine-mapping studies identify risk loci and genetic pathways underlying allergic rhinitis. *Nat. Genet.* **50**, 1072–1080 (2018).
- Muraro, A. et al. Precision medicine in patients with allergic diseases: airway diseases and atopic dermatitis-PRACTALL document of the European Academy of Allergy and Clinical Immunology and the American Academy of Allergy, Asthma & Immunology. *J. Allergy Clin. Immunol.* **137**, 1347–1358 (2016).
- Bougas, N. et al. Unsupervised trajectories of respiratory/allergic symptoms throughout childhood in the PARIS cohort. *Pediatr. Allergy Immunol.* **30**, 315–324 (2019).
- Pinart, M. et al. Comorbidity of eczema, rhinitis, and asthma in IgE-sensitized and non-IgE-sensitized children in MeDALL: a population-based cohort study. *Lancet Respir. Med.* **2**, 131–140 (2014).
The MeDALL study is the first to quantify the net excess of multimorbidity asthma, eczema and AR both in the presence and in the absence of IgE sensitization. It shows that multimorbidity is not occurring by chance.
- Cibella, F. et al. The burden of rhinitis and rhinoconjunctivitis in adolescents. *Allergy Asthma Immunol. Res.* **7**, 44–50 (2015).
- Siroux, V. et al. Association between asthma, rhinitis, and conjunctivitis multimorbidities with molecular IgE sensitization in adults. *Allergy* **74**, 824–827 (2019).
- Toppila-Salmi, S. et al. Risk of adult-onset asthma increases with the number of allergic multimorbidities and decreases with age. *Allergy* <https://doi.org/10.1111/all.13971> (2019).
- Bousquet, J. et al. Daily allergic multimorbidity in rhinitis using mobile technology: a novel concept of the MASK study. *Allergy* **73**, 1622–1631 (2018).
- Amaral, R. et al. Disentangling the heterogeneity of allergic respiratory diseases by latent class analysis reveals novel phenotypes. *Allergy* **74**, 698–708 (2019).
- Jantunen, J. et al. Multimorbidity in asthma, allergic conditions and COPD increase disease severity, drug use and costs: the Finnish pharmacy survey. *Int. Arch. Allergy Immunol.* **179**, 273–280 (2019).
- Bousquet, J. et al. Are allergic multimorbidities and IgE polysensitization associated with the persistence or re-occurrence of foetal type 2 signalling? The MeDALL hypothesis. *Allergy* **70**, 1062–1078 (2015).
- Siroux, V. et al. The asthma-rhinitis multimorbidity is associated with IgE polysensitization in adolescents and adults. *Allergy* **73**, 1447–1458 (2018).
- Pfaar, O. et al. Allergen exposure chambers: harmonizing current concepts and projecting the needs for the future - an EAACI position paper. *Allergy* **72**, 1035–1042 (2017).
- Bousquet, J. et al. Nasal challenge with pollen grains, skin-prick tests and specific IgE in patients with grass pollen allergy. *Clin. Allergy* **17**, 529–536 (1987).
- Durham, S. R. et al. Cytokine messenger RNA expression for IL-3, IL-4, IL-5, and granulocyte/macrophage colony-stimulating factor in the nasal mucosa after local allergen provocation: relationship to tissue eosinophilia. *J. Immunol.* **148**, 2390–2394 (1992).
- Eifan, A. O., Orban, N. T., Jacobson, M. R. & Durham, S. R. Severe persistent allergic rhinitis. Inflammation but no histologic features of structural upper airway remodeling. *Am. J. Respir. Crit. Care Med.* **192**, 1431–1439 (2015).
- Larson, D. et al. Nasal allergen challenge and environmental exposure chamber challenge: a randomized trial comparing clinical and biological responses to cat allergen. *J. Allergy Clin. Immunol.* **145**, 1585–1597 (2020).
- Cameron, L. et al. Local synthesis of epsilon germline gene transcripts, IL-4, and IL-13 in allergic nasal mucosa after ex vivo allergen exposure. *J. Allergy Clin. Immunol.* **106**, 46–52 (2000).
- Lambrecht, B. N. & Hammad, H. Allergens and the airway epithelium response: gateway to allergic sensitization. *J. Allergy Clin. Immunol.* **134**, 499–507 (2014).
- Cayrol, C. et al. Environmental allergens induce allergic inflammation through proteolytic maturation of IL-33. *Nat. Immunol.* **19**, 375–385 (2018).
- Hammad, H. & Lambrecht, B. N. Barrier epithelial cells and the control of type 2 immunity. *Immunity* **43**, 29–40 (2015).
- Roan, F., Obata-Ninomiya, K. & Ziegler, S. F. Epithelial cell-derived cytokines: more than just signaling the alarm. *J. Clin. Invest.* **129**, 1441–1451 (2019).
- Munoz, X. et al. Diesel exhausts particles: their role in increasing the incidence of asthma. Reviewing the evidence of a causal link. *Sci. Total Environ.* **652**, 1129–1138 (2019).
- Schleimer, R. P. & Berdnikovs, S. Etiology of epithelial barrier dysfunction in patients with type 2 inflammatory diseases. *J. Allergy Clin. Immunol.* **139**, 1752–1761 (2017).
- Teufelberger, A. R., Broker, B. M., Krysko, D. V., Bachert, C. & Krysko, O. Staphylococcus aureus orchestrates type 2 airway diseases. *Trends Mol. Med.* **25**, 696–707 (2019).

50. Humbert, M. et al. IgE-mediated multimorbidities in allergic asthma and the potential for omalizumab therapy. *J. Allergy Clin. Immunol. Pract.* **7**, 1418–1429 (2019).
This is a compelling review about the role of IgE in the pathophysiology of allergic asthma and associated allergic multimorbidities, including AR.
51. Palomares, O., Akdis, M., Martin-Fontecha, M. & Akdis, C. A. Mechanisms of immune regulation in allergic diseases: the role of regulatory T and B cells. *Immunol. Rev.* **278**, 219–236 (2017).
This is a comprehensive review discussing the role of regulatory T and B cells in the induction and maintenance of tolerance in allergic diseases, including AR. A deeper analysis of the molecular mechanism governing the generation of regulatory T cells and B cells in tolerance and discussion of how this could be exploited to develop alternative therapeutic interventions is also provided.
52. Palomares, O. et al. dIvERGent: how IgE axis contributes to the continuum of allergic asthma and anti-IgE therapies. *Int. J. Mol. Sci.* **18**, 1328 (2017).
53. Ihara, F. et al. Identification of specifically reduced Th2 cell subsets in allergic rhinitis patients after sublingual immunotherapy. *Allergy* **73**, 1823–1832 (2018).
54. Iinuma, T. et al. Pathogenicity of memory Th2 cells is linked to stage of allergic rhinitis. *Allergy* **73**, 479–489 (2018).
55. Nakayama, T. et al. Th2 cells in health and disease. *Annu. Rev. Immunol.* **35**, 53–84 (2017).
56. Lim, M. C., Taylor, R. M. & Naclerio, R. M. The histology of allergic rhinitis and its comparison to cellular changes in nasal lavage. *Am. J. Respir. Crit. Care Med.* **151**, 136–144 (1995).
57. Renand, A. et al. Synchronous immune alterations mirror clinical response during allergen immunotherapy. *J. Allergy Clin. Immunol.* **141**, 1750–1760 (2018).
58. Eifan, A. O. & Durham, S. R. Pathogenesis of rhinitis. *Clin. Exp. Allergy* **46**, 1139–1151 (2016).
This review illustrates key concepts of the pathogenesis of different forms of rhinitis. Environmental factors in association with imbalance in innate and adaptive immunity factors are likely to play major roles. Evidence for nasal priming and remodelling in AR are reviewed. Local AR in the absence of systemic IgE is discussed. Non-allergic (non-IgE-mediated) rhinitis is considered.
59. Naclerio, R. M. et al. Inflammatory mediators in late antigen-induced rhinitis. *N. Engl. J. Med.* **313**, 65–70 (1985).
60. Connell, J. T. Quantitative intranasal pollen challenges: III. The priming effect in allergic rhinitis. *J. Allergy* **43**, 33–44 (1969).
61. Bousquet, J., Jacquot, W., Vignola, A. M., Bachert, C. & Van Cauwenberge, P. Allergic rhinitis: a disease remodeling the upper airways? *J. Allergy Clin. Immunol.* **113**, 43–49 (2004).
62. Wachs, M. et al. Observations on the pathogenesis of nasal priming. *J. Allergy Clin. Immunol.* **84**, 492–501 (1989).
63. Weisel, F., Shlomchik, M. & Memory, B. Cells of mice and humans. *Annu. Rev. Immunol.* **35**, 255–284 (2017).
64. Schuijs, M. J., Hammad, H. & Lambrecht, B. N. Professional and 'amateur' antigen-presenting cells in type 2 immunity. *Trends Immunol.* **40**, 22–34 (2019).
65. Sarin, S., Undem, B., Sanico, A. & Togias, A. The role of the nervous system in rhinitis. *J. Allergy Clin. Immunol.* **118**, 999–1016 (2006).
66. Chen, J. et al. Expression of bitter taste receptors and solitary chemosensory cell markers in the human sinonasal cavity. *Chem. Senses* **44**, 483–495 (2019).
67. Mosimann, B. L. et al. Substance P, calcitonin gene-related peptide, and vasoactive intestinal peptide increase in nasal secretions after allergen challenge in atopic patients. *J. Allergy Clin. Immunol.* **92**, 95–104 (1993).
68. Undem, B. J. & Taylor-Clark, T. Mechanisms underlying the neuronal-based symptoms of allergy. *J. Allergy Clin. Immunol.* **133**, 1521–1534 (2014).
69. Sanico, A. M. et al. Nerve growth factor expression and release in allergic inflammatory disease of the upper airways. *Am. J. Respir. Crit. Care Med.* **161**, 1631–1635 (2000).
70. Wu, X., Myers, A. C., Goldstone, A. C., Togias, A. & Sanico, A. M. Localization of nerve growth factor and its receptors in the human nasal mucosa. *J. Allergy Clin. Immunol.* **118**, 428–435 (2006).
71. Samolinski, B. et al. Prevention and control of childhood asthma and allergy in the EU from the public health point of view: Polish Presidency of the European Union. *Allergy* **67**, 726–731 (2012).
72. Scadding, G. et al. Diagnostic tools in rhinology EAACI position paper. *Clin. Transl. Allergy* **1**, 2 (2011).
A panel of European experts in the field of rhinology contributed to this consensus document on diagnostic tools in rhinology. This EAACI Task Force document aims at providing readers with a comprehensive and complete overview of currently available tools for diagnosis of nasal and sinonasal disease. The different important issues related to history taking, clinical examination and additional investigative tools for evaluation of the severity of sinonasal disease are logically ordered into a consensus document.
73. Bousquet, J., Van Cauwenberge, P. & Khaltaev, N. Allergic rhinitis and its impact on asthma. *J. Allergy Clin. Immunol.* **108**, S147–S334 (2001).
74. Valero, A. et al. A new criterion by which to discriminate between patients with moderate allergic rhinitis and patients with severe allergic rhinitis based on the allergic rhinitis and its impact on asthma severity items. *J. Allergy Clin. Immunol.* **120**, 359–365 (2007).
75. Bachert, C., Bousquet, J. & Hellings, P. Rapid onset of action and reduced nasal hyperreactivity: new targets in allergic rhinitis management. *Clin. Transl. Allergy* **8**, 25 (2018).
76. Cingi, C. et al. Multi-morbidities of allergic rhinitis in adults: European Academy of Allergy and Clinical Immunology task force report. *Clin. Transl. Allergy* **7**, 17 (2017).
This report prepared by the EAACI Task Force on Allergic Rhinitis Comorbidities aims to highlight the role of multimorbidities in the definition, classification, mechanisms, recommendations for diagnosis and treatment of AR, and to define the needs in this neglected area by a literature review.
77. Skypala, I. J. et al. The prevalence of PFS and prevalence and characteristics of reported food allergy: a survey of UK adults aged 18–75 incorporating a validated PFS diagnostic questionnaire. *Clin. Exp. Allergy* **43**, 928–940 (2013).
78. Mener, D. J., Shargorodsky, J., Varadhan, R. & Lin, S. Y. Topical intranasal corticosteroids and growth velocity in children: a meta-analysis. *Int. Forum Allergy Rhinol.* **5**, 95–103 (2015).
79. Scadding, G. K. et al. BSACI guideline for the diagnosis and management of allergic and non-allergic rhinitis (revised edition 2017; first edition 2007). *Clin. Exp. Allergy* **47**, 856–889 (2017).
80. Lourenco, O. et al. Managing allergic rhinitis in the pharmacy: an ARIA guide for implementation in practice. *Pharmacy* **8**, 85 (2020).
81. Petersen, H. et al. Manifestation of eosinophilic granulomatosis with polyangiitis in head and neck. *Rhinology* **53**, 277–285 (2015).
82. Anstotegui, I. J. et al. IgE allergy diagnostics and other relevant tests in allergy, a World Allergy Organization position paper. *World Allergy Organ. J.* **13**, 100080 (2020).
83. Klimek, L. et al. In-vivo diagnostic test allergens in Europe: a call to action and proposal for recovery plan - an EAACI position paper. *Allergy* <https://doi.org/10.1111/all.14329> (2020).
84. Nevis, I. F., Binkley, K. & Kabali, C. Diagnostic accuracy of skin-prick testing for allergic rhinitis: a systematic review and meta-analysis. *Allergy Asthma Clin. Immunol.* **12**, 20 (2016).
85. Cipriani, F. et al. Early molecular biomarkers predicting the evolution of allergic rhinitis and its comorbidities: a longitudinal multicenter study of a patient cohort. *Pediatr. Allergy Immunol.* **30**, 325–334 (2019).
86. Steering Committee Authors and Review Panel Members. A WAO – ARIA – GA²LEN consensus document on molecular-based allergy diagnosis (PAMD@): update 2020. *World Allergy Organ. J.* **13**, 100091 (2020).
87. Westman, M. et al. Sensitization to grass pollen allergen molecules in a birth cohort-natural Phl p 4 as an early indicator of grass pollen allergy. *J. Allergy Clin. Immunol.* **145**, 1174–1181 (2020).
88. Romero, J. N. & Scadding, G. Eosinophilia in nasal secretions compared to skin prick test and nasal challenge test in the diagnosis of nasal allergy. *Rhinology* **30**, 169–175 (1992).
89. Liu, C. et al. Use of nasal nitric oxide in the diagnosis of allergic rhinitis and nonallergic rhinitis in patients with and without sinus inflammation. *J. Allergy Clin. Immunol. Pract.* **8**, 1574–1581 (2020).
90. Campo, P. et al. Local allergic rhinitis: implications for management. *Clin. Exp. Allergy* **49**, 6–16 (2019).
91. Rondon, C. et al. Local allergic rhinitis is an independent rhinitis phenotype: the results of a 10-year follow-up study. *Allergy* **73**, 470–478 (2018).
92. Raciborski, F. et al. Correction to: dissociating polysensitization and multimorbidity in children and adults from a Polish general population cohort. *Clin. Transl. Allergy* **9**, 23 (2019).
93. Deckers, J., Lambrecht, B. N. & Hammad, H. How a farming environment protects from atopy. *Curr. Opin. Immunol.* **60**, 163–169 (2019).
94. Levin, M. E. et al. Environmental factors associated with allergy in urban and rural children from the South African Food Allergy (SAFFA) cohort. *J. Allergy Clin. Immunol.* **145**, 415–426 (2020).
95. Eller, E. et al. Meta-analysis of determinants for pet ownership in 12 European birth cohorts on asthma and allergies: a GA²LEN initiative. *Allergy* **63**, 1491–1498 (2008).
96. Cuervo-García, C. A. et al. World allergy organization-McMaster university guidelines for allergic disease prevention (GLAD-P): prebiotics. *World Allergy Organ. J.* **9**, 10 (2016).
97. Fiocchi, A. et al. World allergy organization-McMaster university guidelines for allergic disease prevention (GLAD-P): probiotics. *World Allergy Organ. J.* **8**, 4 (2015).
98. Bousquet, J. et al. Prevention of pollen rhinitis symptoms: comparison of fluticasone propionate aqueous nasal spray and disodium cromoglycate aqueous nasal spray. A multicenter, double-blind, double-dummy, parallel-group study. *Allergy* **48**, 327–333 (1993).
99. Scadding, G. K. Optimal management of allergic rhinitis. *Arch. Dis. Child* **100**, 576–582 (2015).
100. Okubo, K., Ogino, S., Nagakura, T. & Ishikawa, T. Omalizumab is effective and safe in the treatment of Japanese cedar pollen-induced seasonal allergic rhinitis. *Allergol. Int.* **55**, 379–386 (2006).
101. Nurmatov, U., van Schayck, C. P., Hurwitz, B. & Sheikh, A. House dust mite avoidance measures for perennial allergic rhinitis: an updated Cochrane systematic review. *Allergy* **67**, 158–165 (2012).
102. Satyaraj, E., Wedner, H. J. & Bousquet, J. Keep the cat, change the care pathway: a transformational approach to managing Fel d 1, the major cat allergen. *Allergy* **74** (Suppl 107), 5–17 (2019).
103. Kawachi, H., Yanai, K., Wang, D. Y., Itahashi, K. & Okubo, K. Antihistamines for allergic rhinitis treatment from the viewpoint of non-sedative properties. *Int. J. Mol. Sci.* **20**, 213 (2019).
104. Brozek, J. L. et al. Allergic rhinitis and its impact on asthma (ARIA) guidelines: 2010 revision. *J. Allergy Clin. Immunol.* **126**, 466–476 (2010).
The GRADE approach is used to propose recommendations for some recent information published on the pharmacological treatment of AR.
105. Church, M. K. et al. Risk of first-generation H₁-antihistamines: a GA²LEN position paper. *Allergy* **65**, 459–466 (2010).
106. Simons, F. E. Advances in H1-antihistamines. *N. Engl. J. Med.* **351**, 2203–2217 (2004).
107. Kar, S., Krishnan, A., Preetha, K. & Mohankar, A. A review of antihistamines used during pregnancy. *J. Pharmacol. Pharmacother.* **3**, 105–108 (2012).
108. Patel, P., D'Andrea, C. & Sacks, H. J. Onset of action of azelastine nasal spray compared with mometasone nasal spray and placebo in subjects with seasonal allergic rhinitis evaluated in an environmental exposure chamber. *Am. J. Rhinol.* **21**, 499–503 (2007).
109. Rodrigo, G. J. & Neffen, H. Efficacy of fluticasone furoate nasal spray vs. placebo for the treatment of ocular and nasal symptoms of allergic rhinitis: a systematic review. *Clin. Exp. Allergy* **41**, 160–170 (2011).
110. Wallace, D. V. & Dykewicz, M. S. Comparing the evidence in allergic rhinitis guidelines. *Curr. Opin. Allergy Clin. Immunol.* **17**, 286–294 (2017).
111. Wu, E. L. et al. Epistaxis risk associated with intranasal corticosteroid sprays: a systematic review and meta-analysis. *Otolaryngol. Head Neck Surg.* **161**, 18–27 (2019).
112. Valenzuela, C. V. et al. Intranasal corticosteroids do not lead to ocular changes: a systematic review and meta-analysis. *Laryngoscope* **129**, 6–12 (2019).
113. Alhussien, A. H., Alhedaithy, R. A. & Alsaleh, S. A. Safety of intranasal corticosteroid sprays during pregnancy: an updated review. *Eur. Arch. Otorhinolaryngol.* **275**, 325–333 (2018).
114. Hampel, F. C. et al. Double-blind, placebo-controlled study of azelastine and fluticasone in a single nasal spray delivery device. *Ann. Allergy Asthma Immunol.* **105**, 168–173 (2010).

115. Segall, N., Prenner, B., Lumry, W., Caracta, C. F. & Tantry, S. K. Long-term safety and efficacy of olopatadine-mometasone combination nasal spray in patients with perennial allergic rhinitis. *Allergy Asthma Proc.* **40**, 301–310 (2019).
116. Bousquet, J. et al. Onset of action of the fixed combination intranasal azelastine-fluticasone propionate in an allergen exposure chamber. *J. Allergy Clin. Immunol. Pract.* **6**, 1726–1732 (2018).
117. Patel, P., Salapatek, A. M. & Tantry, S. K. Effect of olopatadine-mometasone combination nasal spray on seasonal allergic rhinitis symptoms in an environmental exposure chamber study. *Ann. Allergy Asthma Immunol.* **122**, 160–166 (2019).
118. Brozek, J. L. et al. Allergic rhinitis and its impact on asthma (ARIA) guidelines - 2016 revision. *J. Allergy Clin. Immunol.* <https://doi.org/10.1016/j.jaci.2017.03.050> (2017).
119. Wallace, D. V., Dykewicz, M. S., Oppenheimer, J., Portnoy, J. M. & Lang, D. M. Pharmacologic treatment of seasonal allergic rhinitis: synopsis of guidance from the 2017 joint task force on practice parameters. *Ann. Intern. Med.* <https://doi.org/10.7326/M17-2203> (2017).
- The GRADE approach is used to propose recommendations for some recent information published on the pharmacological treatment of AR. For the first time similar US and ARIA recommendations are published.**
120. Hermelingmeier, K. E., Weber, R. K., Hellmich, M., Heubach, C. P. & Mosges, R. Nasal irrigation as an adjunctive treatment in allergic rhinitis: a systematic review and meta-analysis. *Am. J. Rhinol. Allergy* **26**, e119–e125 (2012).
121. Head, K. et al. Saline irrigation for allergic rhinitis. *Cochrane Database Syst. Rev.* **6**, CD012597 (2018).
122. Kozlov, V., Lavrenova, G., Savlevich, E. & Bazarkina, K. Evidence-based phytotherapy in allergic rhinitis. *Clin. Phytosci.* **4**, 23 (2018).
123. Mullol, J. et al. Update on rupatadine in the management of allergic disorders. *Allergy* **70** (Suppl 100), 1–24 (2015).
124. Nayak, A. S. et al. Randomized, placebo-controlled study of cetirizine and loratadine in children with seasonal allergic rhinitis. *Allergy Asthma Proc.* **38**, 222–230 (2017).
125. Berger, W. et al. Efficacy of MP-AzeFlu in children with seasonal allergic rhinitis: importance of paediatric symptom assessment. *Pediatr. Allergy Immunol.* **27**, 126–133 (2016).
126. Settipane, R. A. & Kaliner, M. A. Chapter 14: nonallergic rhinitis. *Am. J. Rhinol. Allergy* **27**, 48–51 (2013).
127. Bozek, A. Pharmacological management of allergic rhinitis in the elderly. *Drugs Aging* **34**, 21–28 (2017).
128. Kaliner, M. A. H1-antihistamines in the elderly. *Clin. Allergy Immunol.* **17**, 465–481 (2002).
129. Davila, I. et al. Use of second generation H1 antihistamines in special situations. *J. Investig. Allergol Clin. Immunol.* **23** (Suppl 1), 1–16 (2013).
130. Bosnic-Anticevich, S. et al. ARIA pharmacy 2018 “Allergic rhinitis care pathways for community pharmacy”: AIRWAYS ICPs initiative (European Innovation Partnership on Active and Healthy Ageing, DG CONNECT and DG Sante) POLLAR (Impact of air pollution on asthma and rhinitis) GARD Demonstration project. *Allergy* **74**, 1219–1236 (2019).
- Guidance for pharmacists to manage AR is proposed.**
131. Menditto, E. et al. Adherence to treatment in allergic rhinitis using mobile technology. *The MASK study. Clin. Exp. Allergy* **49**, 442–460 (2019).
132. Bedard, A. et al. Mobile technology offers novel insights into the control and treatment of allergic rhinitis: the MASK study. *J. Allergy Clin. Immunol.* **144**, 135–143 (2019).
133. Brozek, J. L. et al. Methodology for development of the allergic rhinitis and its impact on asthma guideline 2008 update. *Allergy* **63**, 38–46 (2008).
134. Bousquet, J. et al. Next-generation Allergic Rhinitis and its Impact on Asthma (ARIA) guidelines for allergic rhinitis based on Grading of Recommendations Assessment, Development and Evaluation (GRADE) and real-world evidence. *J. Allergy Clin. Immunol.* **145**, 70–80 e73 (2020).
- This is the first report for any disease combining evidence-based medicine using the GRADE approach and real-world evidence (real-world data using mobile technology and chamber studies to assess the speed of onset of medications). Next-generation guidelines differ from the GRADE recommendations.**
135. Bousquet, J. et al. Guidance to 2018 good practice: ARIA digitally-enabled, integrated, person-centred care for rhinitis and asthma. *Clin. Transl. Allergy* **9**, 16 (2019).
136. Bousquet, J. et al. From ARIA guidelines to the digital transformation of health in rhinitis and asthma multimorbidity. *Eur. Respir. J.* <https://doi.org/10.1183/13993003.01023-2019> (2019).
137. Bousquet, J. J. et al. Next-generation ARIA care pathways for rhinitis and asthma: a model for multimorbid chronic diseases. *Clin. Transl. Allergy* **9**, 44 (2019).
138. Bousquet, J. et al. 2019 ARIA care pathways for allergen immunotherapy. *Allergy* **74**, 2087–2102 (2019).
139. Penagos, M. & Durham, S. R. Duration of allergen immunotherapy for inhaled allergy. *Curr. Opin. Allergy Clin. Immunol.* **19**, 594–605 (2019).
140. Sharif, H. et al. Immunologic mechanisms of a short-course of Lolium perenne peptide immunotherapy: a randomized, double-blind, placebo-controlled trial. *J. Allergy Clin. Immunol.* <https://doi.org/10.1016/j.jaci.2019.02.023> (2019).
141. Shamji, M. H. et al. Nasal allergen-neutralizing IgG4 antibodies block IgE-mediated responses: novel biomarker of subcutaneous grass pollen immunotherapy. *J. Allergy Clin. Immunol.* **143**, 1067–1076 (2019).
142. Zielen, S., Devillier, P., Heinrich, J., Richter, H. & Wahn, U. Sublingual immunotherapy provides long-term relief in allergic rhinitis and reduces the risk of asthma: a retrospective, real-world database analysis. *Allergy* **73**, 165–177 (2018).
143. Okamoto, Y. et al. Efficacy and safety of sublingual immunotherapy for two seasons in patients with Japanese cedar pollinosis. *Int. Arch. Allergy Immunol.* **166**, 177–188 (2015).
144. Gotoh, M. et al. Long-term efficacy and dose-finding trial of Japanese cedar pollen sublingual immunotherapy tablet. *J. Allergy Clin. Immunol. Pract.* **7**, 1287–1297 (2019).
145. Virchow, J. C. et al. Efficacy of a house dust mite sublingual allergen immunotherapy tablet in adults with allergic asthma: a randomized clinical trial. *JAMA* **315**, 1715–1725 (2016).
146. Bachert, C. et al. Allergen immunotherapy on the way to product-based evaluation: a WAO statement. *World Allergy Organ J.* **8**, 29 (2015).
147. Bonertz, A., Mahler, V. & Vieths, S. Manufacturing and quality assessment of allergenic extracts for immunotherapy: state of the art. *Curr. Opin. Allergy Clin. Immunol.* **19**, 640–645 (2019).
148. Pitsios, C. et al. Contraindications to immunotherapy: a global approach. *Clin. Transl. Allergy* **9**, 45 (2019).
149. Pfaar, O. et al. Guideline on allergen-specific immunotherapy in IgE-mediated allergic diseases: S2k guideline of the German Society for Allergy and Clinical Immunology (DGAKI), the Society for Pediatric Allergy and Environmental Medicine (GPA), the Medical Association of German Allergologists (AeDA), the Austrian Society for Allergy and Immunology (OGAI), the Swiss Society for Allergy and Immunology (SGAI), the German Society of Dermatology (DDG), the German Society of Oto-Rhino-Laryngology, Head and Neck Surgery (DGHN0-KHC), the German Society of Pediatrics and Adolescent Medicine (DGKJ), the Society for Pediatric Pneumology (GPP), the German Respiratory Society (DGP), the German Association of ENT Surgeons (BV-HNO), the Professional Federation of Paediatricians and Youth Doctors (BVKJ), the Federal Association of Pulmonologists (BDP) and the German Dermatologists Association (BVDD). *Allergo J. Int.* **23**, 282–319 (2014).
150. Muraro, A. et al. EAACI guidelines on allergen immunotherapy: executive statement. *Allergy* **73**, 739–743 (2018).
151. Roberts, G. et al. EAACI guidelines on allergen immunotherapy: allergic rhinoconjunctivitis. *Allergy* **73**, 765–798 (2018).
152. Agache, I. et al. EAACI guidelines on allergen immunotherapy: house dust mite-driven allergic asthma. *Allergy* **74**, 855–873 (2019).
153. Shamji, M. H. & Durham, S. R. Mechanisms of allergen immunotherapy for inhaled allergens and predictive biomarkers. *J. Allergy Clin. Immunol.* **140**, 1485–1498 (2017).
154. US Department of Health and Human Services FDA Center for Drug Evaluation and Research, US Department of Health and Human Services FDA Center for Biologics Evaluation and Research & US Department of Health and Human Services FDA Center for Devices and Radiological Health. Guidance for Industry: patient reported outcome measures: use in medical product development to support labeling claims: draft guidance. *Health Qual. Life Outcomes* **4**, 79 (2006).
155. Committee for medicinal products for human use (CHMP). Reflection paper on the regulatory guidance for the use of health-related quality of life (HRQL) measures in the evaluation of medicinal products. *European Medicines Agency* https://www.ema.europa.eu/en/documents/scientific-guideline/reflection-paper-regulatory-guidance-use-healthrelated-quality-life-hrql-measures-evaluation_en.pdf (2005).
156. Brozek, J. L. et al. Grading quality of evidence and strength of recommendations in clinical practice guidelines. Part 1 of 3. An overview of the GRADE approach and grading quality of evidence about interventions. *Allergy* **64**, 669–677 (2009).
157. Sprangers, M. A., Hall, P., Morisky, D. E., Narrow, W. E. & Dapuerto, J. Using patient-reported measurement to pave the path towards personalized medicine. *Qual. Life Res.* **22**, 2631–2637 (2013).
158. Leynaert, B., Neukirch, C., Liard, R., Bousquet, J. & Neukirch, F. Quality of life in allergic rhinitis and asthma. A population-based study of young adults. *Am. J. Respir. Crit. Care Med.* **162**, 1391–1396 (2000).
159. Sritipsukho, P. & Viriyaudomsri, O. Health-related quality of life in Thai children with allergic respiratory diseases. *J. Med. Assoc. Thai.* **98**, 457–463 (2015).
160. Sritipsukho, P., Satdhabudha, A. & Nanthapais, S. Effect of allergic rhinitis and asthma on the quality of life in young Thai adolescents. *Asian Pac. J. Allergy Immunol.* **33**, 222–226 (2015).
161. Dietz de Loos, D. A., Segboer, C. L., Gevorgyan, A. & Fokkens, W. J. Disease-specific quality-of-life questionnaires in rhinitis and rhinosinusitis: review and evaluation. *Curr. Allergy Asthma Rep.* **13**, 162–170 (2013).
162. Calderon, M. A., Casale, T. B. & Demoly, P. Validation of patient-reported outcomes for clinical trials in allergic rhinitis: a systematic review. *J. Allergy Clin. Immunol. Pract.* **7**, 1450–1461 (2019).
163. Meltzer, E. O. Allergic rhinitis: burden of illness, quality of life, comorbidities, and control. *Immunol. Allergy Clin. North Am.* **36**, 235–248 (2016).
164. Braidó, F. et al. Patients with asthma and comorbid allergic rhinitis: is optimal quality of life achievable in real life? *PLoS ONE* **7**, e31178 (2012).
165. Baiardini, I. et al. RHINASTHMA-children: a new quality of life tool for patients with respiratory allergy. *Pediatr. Allergy Immunol.* **28**, 102–105 (2017).
166. La Grutta, S. et al. RHINASTHMA-adolescents: a new quality of life tool for patients with respiratory allergy. *Pediatr. Allergy Immunol.* **25**, 450–455 (2014).
167. Ilyina, N. I. et al. Efficacy of a novel intranasal formulation of azelastine hydrochloride and fluticasone propionate, delivered in a single spray, for the treatment of seasonal allergic rhinitis: results from Russia. *Int. Arch. Allergy Immunol.* **178**, 255–263 (2019).
168. Lavallee, D. C. et al. Incorporating patient-reported outcomes into health care to engage patients and enhance care. *Health Aff.* **35**, 575–582 (2016).
169. Harle, C. A. et al. Overcoming barriers to implementing patient-reported outcomes in an electronic health record: a case report. *J. Am. Med. Inform. Assoc.* **23**, 74–79 (2016).
170. Braidó, F. et al. RhinAsthma patient perspective: a short daily asthma and rhinitis QoL assessment. *Allergy* **67**, 1443–1450 (2012).
171. Matricardi, P. M. et al. The role of mobile health technologies in allergy care: an EAACI position paper. *Allergy* **75**, 259–272 (2020).
172. Tripodi, S. et al. Digital technologies for an improved management of respiratory allergic diseases: 10 years of clinical studies using an online platform for patients and physicians. *Ital. J. Pediatr.* **46**, 105 (2020).
173. Bousquet, J. et al. Mobile technology in allergic rhinitis: evolution in management or revolution in health and care? *J. Allergy Clin. Immunol. Pract.* **7**, 2511–2523 (2019).
174. Basti, K., Berger, U. & Kmenta, M. Evaluation of pollen apps forecasts: the need for quality control in an eHealth service. *J. Med. Internet Res.* **19**, e152 (2017).
175. Sofiev, M. et al. A demonstration project of global alliance against chronic respiratory diseases: prediction of interactions between air pollution and allergen exposure—the Mobile Airways Sentinel Network-Impact of air pollution on asthma and rhinitis approach. *Chin. Med. J.* <https://doi.org/10.1097/CM9.0000000000000916> (2020).

176. Jiang, Y. et al. Transcriptomics of atopy and atopic asthma in white blood cells from children and adolescents. *Eur. Respir. J.* <https://doi.org/10.1183/13993003.00102-2019> (2019).
177. Aguilar, D. et al. Understanding allergic multimorbidity within the non-eosinophilic interactome. *PLoS ONE* **14**, e0224448 (2019).
178. Heffler, E. et al. New drugs in early-stage clinical trials for allergic rhinitis. *Expert Opin. Investig. Drugs* **28**, 267–273 (2019).
179. Yu, C. et al. Clinical efficacy and safety of omalizumab in the treatment of allergic rhinitis: a systematic review and meta-analysis of randomized clinical trials. *Am. J. Rhinol. Allergy* **34**, 196–208 (2020).
180. Patel, G. B., Kern, R. C., Bernstein, J. A., Hae-Sim, P. & Peters, A. T. Current and future treatments of rhinitis and sinusitis. *J. Allergy Clin. Immunol. Pract.* <https://doi.org/10.1016/j.jaip.2020.01.031> (2020).
181. Pfaar, O. et al. Perspectives in allergen immunotherapy: 2019 and beyond. *Allergy* **74** (Suppl 108), 3–25 (2019).
182. Dorofeeva, Y. et al. Past, presence and future of allergen immunotherapy vaccines. *Allergy* <https://doi.org/10.1111/all.14300> (2020).
183. Pechsrichuang, P. & Jacquet, A. Molecular approaches to allergen-specific immunotherapy: are we so far from clinical implementation? *Clin. Exp. Allergy* <https://doi.org/10.1111/cea.13588> (2020).
184. Rodriguez-Dominguez, A. et al. Molecular profiling of allergen-specific antibody responses may enhance success of specific immunotherapy. *J. Allergy Clin. Immunol.* <https://doi.org/10.1016/j.jaci.2020.03.029> (2020).
185. Komlosi, Z. I. et al. Highlights of novel vaccination strategies in allergen immunotherapy. *Immunol. Allergy Clin. North Am.* **40**, 15–24 (2020).
186. Massanari, M. et al. Effect of pretreatment with omalizumab on the tolerability of specific immunotherapy in allergic asthma. *J. Allergy Clin. Immunol.* **125**, 383–389 (2010).
187. Nelson, H. S. Allergy immunotherapy: future directions for the 2020s. *Allergy Asthma Proc.* **41**, 314–325 (2020).
188. Tulaeva, I. et al. Preventive allergen-specific vaccination against allergy: mission possible? *Front. Immunol.* **11**, 1368 (2020).
189. Toumi, M. & Remuzat, C. Value added medicines: what value repurposed medicines might bring to society? *J. Mark Access Health Policy* **5**, 1264717 (2017).
190. Bousquet, J. et al. ARIA digital anamorphosis: digital transformation of health and care in airway diseases from research to practice. *Allergy* <https://doi.org/10.1111/all.14422> (2020).
191. Stenberg, K., Lauer, J. A., Kkontouras, G., Fitzpatrick, C. & Stanciole, A. Econometric estimation of WHO-CHOICE country-specific costs for inpatient and outpatient health service delivery. *Cost Eff. Resour. Alloc.* **16**, 11 (2018).
192. Russell, J. & Greenhalgh, T. Affordability as a discursive accomplishment in a changing National Health Service. *Soc. Sci. Med.* **75**, 2463–2471 (2012).
193. Hunter, D. J. et al. Doing transformational change in the English NHS in the context of “big bang” reorganisation. *J. Health Organ. Manag.* **29**, 10–24 (2015).
194. Farmanova, E., Bonneville, L. & Bouchard, L. Organizational health literacy: review of theories, frameworks, guides, and implementation issues. *Inquiry* **55**, 46958018757848 (2018).
195. Bousquet, J. et al. Allergic Rhinitis and its Impact on Asthma (ARIA) phase 4 (2018): change management in allergic rhinitis and asthma multimorbidity using mobile technology. *J. Allergy Clin. Immunol.* **143**, 864–879 (2019).
196. Juniper, E. F. & Guyatt, G. H. Development and testing of a new measure of health status for clinical trials in rhinoconjunctivitis. *Clin. Exp. Allergy* **21**, 77–83 (1991).
197. Juniper, E. F., Thompson, A. K., Ferrie, P. J. & Roberts, J. N. Development and validation of the mini rhinoconjunctivitis quality of life questionnaire. *Clin. Exp. Allergy* **30**, 132–140 (2000).
198. Juniper, E. F., Rohrbach, T. & Meltzer, E. O. A questionnaire to measure quality of life in adults with nocturnal allergic rhinoconjunctivitis. *J. Allergy Clin. Immunol.* **111**, 484–490 (2003).
199. Baiardini, I. et al. Rhinasthma: a new specific QoL questionnaire for patients with rhinitis and asthma. *Allergy* **58**, 289–294 (2003).
200. Juniper, E. F., Guyatt, G. H. & Dolovich, J. Assessment of quality of life in adolescents with allergic rhinoconjunctivitis: development and testing of a questionnaire for clinical trials. *J. Allergy Clin. Immunol.* **93**, 413–423 (1994).
201. Juniper, E. F., Howland, W. C., Roberts, N. B., Thompson, A. K. & King, D. R. Measuring quality of life in children with rhinoconjunctivitis. *J. Allergy Clin. Immunol.* **101**, 163–170 (1998).
202. Fasola, S. et al. RAPP-children: a new tool for assessing quality of life in patients with asthma and rhinitis. *Clin. Exp. Allergy* <https://doi.org/10.1111/cea.13599> (2020).
203. Yuki, K., Fujiogi, M. & Koutsogiannaki, S. COVID-19 pathophysiology: a review. *Clin. Immunol.* **215**, 108427 (2020).
204. Jackson, D. et al. Association of respiratory allergy, asthma and expression of the SARS-CoV-2 receptor, ACE2. *J. Allergy Clin. Immunol.* **146**, 203–206 (2020).
205. Kimura, H. et al. Type 2 inflammation modulates ACE2 and TMPRSS2 in airway epithelial cells. *J. Allergy Clin. Immunol.* <https://doi.org/10.1016/j.jaci.2020.05.004> (2020).
206. Yang, J. M. et al. Allergic disorders and susceptibility to and severity of COVID-19: a nationwide cohort study. *J. Allergy Clin. Immunol.* <https://doi.org/10.1016/j.jaci.2020.08.008> (2020).
207. Lechien, J. R. et al. Olfactory and gustatory dysfunctions as a clinical presentation of mild-to-moderate forms of the coronavirus disease (COVID-19): a multicenter European study. *Eur. Arch. Otorhinolaryngol.* <https://doi.org/10.1007/s00405-020-05965-1> (2020).
208. Pfaar, O. et al. COVID-19 pandemic: practical considerations on the organization of an allergic clinic – an EAACI/ARIA position paper. *Allergy* <https://doi.org/10.1111/all.14453> (2020).
209. Radulesco, T. et al. COVID-19 and rhinology, from the consultation room to the operating theatre. *Eur. Ann. Otorhinolaryngol. Head Neck Dis.* <https://doi.org/10.1016/j.anorl.2020.04.013> (2020).
210. Riggioni, C. et al. A compendium answering 150 questions on COVID-19 and SARS-CoV-2. *Allergy* <https://doi.org/10.1111/all.14449> (2020).
211. Zhang, Y. & Zhang, L. Management practice of allergic rhinitis in china during the COVID-19 pandemic. *Allergy Asthma Immunol. Res.* **12**, 738–742 (2020).
212. Codispoli, C. D., Bandi, S., Moy, J. & Mahdavinia, M. Running a virtual allergy division and training program in the time of COVID-19 pandemic. *J. Allergy Clin. Immunol.* <https://doi.org/10.1016/j.jaci.2020.03.018> (2020).
213. Bousquet, J. et al. Intranasal corticosteroids in allergic rhinitis in COVID-19 infected patients: an ARIA-EAACI statement. *Allergy* <https://doi.org/10.1111/all.14302> (2020).
214. Klimek, L. et al. Allergen immunotherapy in the current COVID-19 pandemic: a position paper of AeDA, ARIA, EAACI, DGAKI and GPA: position paper of the German ARIA Group^a in cooperation with the Austrian ARIA Group^b, the Swiss ARIA Group^c, German Society for Applied Allergy (AEDA)^d, German Society for Allergy and Clinical Immunology (DGAKI)^e, Society for Pediatric Allergy (GPA)^f in cooperation with AG Clinical Immunology, Allergy and Environmental Medicine of the DGHNO-KHC^g and the European Academy of Allergy and Clinical Immunology (EAACI)^h. *Allergol. Select.* **4**, 44–52 (2020).
215. Klimek, L. et al. Handling of allergen immunotherapy in the COVID-19 pandemic: an ARIA-EAACI statement. *Allergy* <https://doi.org/10.1111/all.14336> (2020).
216. Malpiero, G. et al. An academic allergy unit during COVID-19 pandemic in Italy. *J. Allergy Clin. Immunol.* <https://doi.org/10.1016/j.jaci.2020.04.003> (2020).
217. Dror, A. A. et al. Reduction of allergic rhinitis symptoms with face mask usage during the COVID-19 pandemic. *J. Allergy Clin. Immunol. Pract.* <https://doi.org/10.1016/j.jaip.2020.08.035> (2020).

Author contributions

Introduction (G.W.C.); Epidemiology (J.M.A.); Mechanisms/pathophysiology (J.B., A.T. and O.P.); Genetics (E.M.); Diagnosis, screening and prevention (G.K.S.); Management (S.T.-S., S.B.-A. and C.B.); Quality of life (G.W.C. and I.B.); Outlook (J.B.); Overview of Primer (J.B.).

Competing interests

J.B. reports personal fees from Chiesi, Cipla, Hikma, Menarini, Mundipharma, Mylan, Novartis, Purina, Sanofi-Aventis, Takeda, Teva and Uriach and other fees from KYomed-Innov and MASK-air outside the submitted work. S.B.-A. reports grants from TEVA and personal fees from AstraZeneca, Boehringer Ingelheim, GlaxoSmithKline, Mylan, Sanofi and TEVA outside the submitted work. C.B. reports personal fees from ALK, Mylan, GlaxoSmithKline, Sanofi, Novartis and AstraZeneca. G.W.C. reports having received research grants as well as being a lecturer or having received advisory board fees from A.Menarini, ALK-Abelló, Allergy Therapeutics, AstraZeneca, Boehringer Ingelheim, Chiesi Farmaceutici, Genentech, Guidotti-Malesci, GlaxoSmithKline, Hal Allergy, Mylan, Merck, Merck Sharp & Dohme Mundipharma, Novartis, Regeneron, Roche, Sanofi-Aventis, Sanofi-Genzyme, Stallergenes-Greer, UCB Pharma, Uriach Pharma, Valeas and Vibor-Pharma. I.B. reports personal fees from Menarini, Mundipharma, Novartis, GlaxoSmithKline, Sanofi-Genzyme, AstraZeneca and Boehringer Ingelheim outside the submitted work. E.M. reports personal fees from AstraZeneca, Chiesi, Novartis and Sanofi outside the submitted work. O.P. reports research grants from Immunotek S.L. and Novartis and fees for giving scientific lectures from Allergy Therapeutics, Amgen, AstraZeneca, Diater, GlaxoSmithKline S.A., Immunotek S.L., Novartis, Sanofi-Genzyme and Stallergenes, and has participated in advisory boards for Novartis and Sanofi-Genzyme. G.K.S. reports personal fees from ALK, Bayer, GlaxoSmithKline and Mylan, outside the submitted work, and is British Society for Allergy and Clinical Immunology Rhinitis Guidelines Chair and European Forum for Research and Education in Allergy and Airway Diseases (EUFOREA) Lead for allergic rhinitis. All other authors declare no competing interests.

Disclaimer

A.T.'s co-authorship of this publication does not constitute endorsement by the US National Institute of Allergy and Infectious Diseases, the US National Institutes of Health or any other agency of the US government.

Peer review information

Nature Reviews Disease Primers thanks F. Baroody, P. Devillier, C. Incovaia, Y. Okamoto and the other, anonymous, reviewer(s) for their contribution to the peer review of this work.

Publisher's note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

© Springer Nature Limited 2020