

ORIGINAL ARTICLE

Monoterpene emissions from three Nothofagus species in Patagonia, Argentina

Mauro Centritto^a*, Carlos M. Di Bella^b, Rita Baraldi^c, Maria Eugenia Beget^b, Alejandra Kemerer^b, Francesca Rapparini^c, Patricio Oricchio^b, Cesar Rebella^b and Francesco Loreto^a

^aIstituto di Biologia Agroambientale e Forestale, Consiglio Nazionale delle Ricerche, Monterotondo Scalo (Roma), Italy; ^bInstituto de Clima y Agua, INTA-Castelar, Castelar, Buenos Aires, Argentina; ^cIstituto di Biometeorologia, Sezione di Bologna, Consiglio Nazionale delle Ricerche, Bologna, Italy

(Received 2 November 2007; final version received 12 December 2007)

Isoprenoid emissions have key roles in plant biology and plant interactions with the environment. Global emission inventories of isoprenoid emissions still lack information from a large number species, especially from South American vegetation other than the rainforest ecosystem. A study was conducted to identify the basal emission of isoprenoid under field conditions from three *Nothofagus* species. The three *Nothofagus* species were characterized as strong monoterpene emitters while the emission of isoprene was undetectable. The two deciduous species, *N. pumilio* and *N. antarctica*, had similar photosynthetic parameters, but monoterpene emission rate and, consequently, the fraction of photosynthetic carbon re-emitted in the atmosphere as monoterpenes, were more than three-fold higher in *N. pumilio* than in *N. Antarctica*. The evergreen species *N. dombeyi* showed intermediate values of both monoterpene emission rate and fraction of photosynthetic carbon re-emitted. The monoterpene emission spectrum was very similar among the three *Nothofagus* species screened, but clearly different from the spectrum reported in other monoterpene-emitting species of the Fagaceae family. The importance of these findings for atmospheric chemistry and phylogenic evolution are discussed.

Keywords: emission inventory; gas exchange; isoprenoids; limonene; photosynthesis

Introduction

All plants emit a wide range of volatile compounds among which biogenic volatile organic compounds (BVOCs), often referred also as non-methane organic compounds, have received particular attention in the last two decades (Guenther et al. 1993; Centritto et al. 2004; Brilli et al. 2007; Grote and Niinemets in press). Isoprenoids (isoprene, monoterpenes and sesquiterpenes) make the largest fraction of BVOC emissions. They were discovered to enhance and modulate plant tolerance to heat, pollutants, oxidative stress and abiotic stresses (Sharkey and Yeh 2001; Niinemets et al. 2004), and to affect plant-plant and plant-insect interactions (Gershenzon and Dudareva 2007). The global carbon emitted as BVOCs is about 1.1 Pg per year, half of which is emitted as isoprene, and is believed to be of the same order of magnitude than methane emissions (Guenther et al. 2000). The importance of isoprenoids also lies in their reactivity and potential for profoundly influencing the chemical and physical properties of the atmosphere (Kavouras et al. 1998; Di Carlo et al. 2004). In the atmosphere, isoprenoids rapidly react with hydroxyl radical (OH), the primary oxidizing agent of the atmosphere, affecting its tropospheric concentration (Atkinson and Arey 2003; Di Carlo et al. 2004). In non-polluted areas, isoprenoids cause the depletion of OH radicals in the troposphere and increase the lifetimes of other greenhouse gases (including methane) (Poisson et al. 2000). In polluted areas, in the presence of high NO_X concentrations, isoprenoids react in the atmosphere with anthropogenic compounds leading to a net production of tropospheric ozone and other photo-oxidants (Chameides et al. 1987; Fehsenfeld et al. 1992).

Furthermore, monoterpene and sesquiterpenes are significant precursors of secondary organic aerosols (Andreae and Crutzen 1997; Kavouras et al. 1998; Kanakidou et al. 2004; Vizuete et al. 2004). These less-volatile BVOC species condense onto particles in the atmosphere, resulting in the formation of secondary organic aerosols (SOA) which accounts for 20–50% of the total fine particulate matter in the atmosphere at continental mid-latitudes and up to 90% in tropical forested areas (Andreae and Crutzen 1997; Kanakidou et al. 2004). Moreover, although isoprenoid emission accounts for approximately 2% of the total C-exchange of 69 Pg between the biota and the atmosphere (Lal 1999), isoprenoids have not been considered in global C-cycling so far. This emphasizes the importance of biogenic emissions, and inventories of BVOC emissions are, consequently, a key issue in atmospheric sciences.

The emission of isoprenoids by plants was first discovered in the Amazons (Rasmussen and Jones 1973). Afterwards, inventories of the emission of

ISSN 1742-9145 print/ISSN 1742-9153 online © 2008 Taylor & Francis DOI: 10.1080/17429140701861735

http://www.informaworld.com

^{*}Corresponding author. Email: mauro.centritto@ibaf.cnr.it.

isoprenoids have been made systematically for a large number of species in Europe (Simpson et al. 1995; Niinemets et al. 2002), North America (Guenther et al. 1996a, 2000), in Africa (Guenther et al. 1996b), Asia (Loreto et al. 2002; Geron et al. 2006), Australia (He et al. 2000), and South America (Kahl et al. 1999; Rinne et al. 2002; Greenberg et al. 2004). However, inventories about South American plant species are mostly limited to the rainforest vegetation over the Amazon, while information for other ecosystems is missing. More generally, the emission potential databases still lack information especially for monoterpene emitting species, which complicates the use of the emission inventories for predicting the emissions in globally changing environmental conditions (Grote and Niinemets in press). In the past, studies concerning the occurrence of isoprenoid production within the plant kingdom showed that many broad-leaved species were 'non-emitters' or low monoterpene emitters, in particular the species of the Fagaceae family (i.e. Fagus, Nothofagus, Chrysolepis, Castanea and Lithocarpus) (Kesselmeier and Staudt 1999). However, recent studies showed that Fagus sylvatica (European beech) is a moderately strong monoterpene emitter (Dindorf et al. 2006; Holzke et al. 2006).

The current series of experiments was designed to study emissions from temperate forests species of Argentina. We focused on three Nothofagus species (N. antarctica, N. pumilio and N. dombeyi) growing in natural forests in Patagonia, for the following reasons: (a) the Nothofagus genus (southern beech) is important and widespread being formed by 40 species of woody native plants of the southern hemisphere; (b) to our knowledge these three *Nothofagus* species have not been previously screened for BVOC emission; and (c) other species of the Fagaceae family (i.e. oaks) were reported to be strong emitters of isoprene or monoterpenes worldwide in the northern hemisphere (Loreto 2002). However, Fagus sylvatica, the closest plant species to Nothofagus among those vegetating in the northern hemisphere (Cronquist 1981), is controversial with respect to the emission of isoprenoids (Hewitt and Street 1992; Kesselmeier and Staudt 1999; Dindorf et al. 2006).

Materials and methods

Fully sunlit plants of *N. antarctica*, *N. pumilio* and *N. dombeyi* growing in natural forests in San Carlos de Bariloche (Argentina, Lat. 41.07°S, Long. 71.19°W), were studied during late summer 2005. *N. antarctica* and *N. pumilio* are deciduous species, whereas *N. dombeyi* is an evergreen species.

All gas exchange measurements were made between 11:00 and 15:00 h on the central section of a newly-expanded leaf from sunny branches collected early in the morning from 12–15 trees per species and placed in individual 0.5 dm³ containers. Leaves within the same species were at similar phenological stage, without interference from water stress (i.e. relative water content values ranging between 90 and 95%, data not shown). Each branch was cut again under water before measuring gas exchange. The branches were transported to a near laboratory and remained under water from the time of being cut until the measurements were completed. It should be noted that gas exchange parameters measured in these branches equaled rates measured *in situ* (i.e. in native atmospheric conditions), indicating that this procedure did not cause any loss in photosynthetic potential. These gas exchange measurements were made in ambient CO2 concentration with a portable infrared analyser (Licor 6400, Li-Cor, Lincoln, Nebraska, USA).

To enable measurements of photosynthetic basal isoprenoid emission rates, leaves were illuminated using a red-blue light source attached to the gasexchange system which maintain a PPFD (photosynthetic photon flux density) of 1000 μ mol m⁻²s⁻¹. The leaf temperature was controlled with Peltier thermoresistances and was maintained at 30°C. These conditions were selected because they have been reported as the environmental conditions at which the basal emission of isoprenoids is measured (Guenther et al. 1993). The vapor pressure deficit was maintained below 30 mbar bar^{-1} and the relative humidity in the cuvette was about 45%. The leaf was exposed to a flux of 0.51 min^{-1} of ambient air. When CO₂ uptake and H₂O release had become steady, photosynthesis (A), transpiration, stomatal conductance (g_s) were measured and the air exiting the cuvette was diverted from the gas-exchange system to a two-stage trap filled with Carbograph 1 (0.034 g)and Carbograph 2 (0.17 g; Lara, Rome, Italy). The air flowing through the cartridge was measured by a flow meter placed at the cartridge exit. Before each measurement, ambient air from an empty IRGA cuvette was flown through the cartridge to determine the blank. After sampling 2-3 l of air the cartridge was removed, tightly capped, and maintained at low temperature and in the dark until analytical measurement on its content was carried out in Italy. Isoprenoids were analyzed by gas chromatography mass spectrometry as previously described by Baraldi et al. (1999). The trapped compounds were thermodesorbed at 250°C and cryofocused at -150°C on a fused silica liner using a Thermal Desorption Cold Trap Injector (Chrompack, Middleburg, The Netherlands) connected to a gas chromatograph-mass spectrometer (Hewlett Packard 5890-5970, Palo Alto, CA, USA). The desorbed sample was injected into a 60 m \times 0.25 mm I.D. 0.25 µm film thickness capillary column (HP1, Hewlett Packard). Isoprenoids were separated maintaining the oven temperature at 40°C for 10 min and programming to 220°C at 5° C min⁻¹. The identity of the compounds of interest was achieved by comparison of their retention time and mass spectra to that of authentic standards. Quantification of isoprenoids was performed after calculation of standard curves and response factors for each compound, and using d14-cymene as internal standards. Isoprenoid concentration was calculated by dividing the amount collected in the cartridge by the volume of air sampled. Then isoprenoid emissions were calculated by multiplying the difference between isoprenoid concentrations in the sampling and in the corresponding blank by the air flowing through the leaf cuvette. Isoprenoid emission was then referred to leaf area by dividing the emission for the area of the leaf enclosed in the cuvette.

Data were tested using a simple factorial AN-OVA, and where appropriate, the treatment means of gas exchange parameters were compared using Tukey's Post-hoc test.

Results and discussion

There were inherently different gas-exchange characteristics between the two deciduous species and the evergreen species of Nothofagus measured (Table 1). Photosynthesis, stomatal conductance and respiration in darkness were significantly higher in N. antarctica and N. pumilio (the two deciduous species) than in N. dombeyi (the evergreen species). Photosynthesis was on average $\sim 31\%$ higher in the deciduous species than in the evergreen one. Whereas g_s was ~58% and R_D ~40% higher on average in N. antarctica and N. pumilio than in N. dombevi. These results are in keeping with previous findings showing that potential carbon gain (photosynthesis) and carbon loss (respiration) increase in similar proportion with decreasing leaf life-span, increasing leaf nitrogen concentration, and increasing leaf surface area-to-mass ratio (Reich et al. 1997; Wright et al. 2004).

Basal emissions of isoprenoids were measured at photosynthetic photon flux density of 1000 µmol $m^{-2}s^{-1}$ and at a leaf temperature of 30°C to normalize the emission dependency on environmental factors (Guenther et al. 2000) in fully expanded leaves of adult trees. All *Nothofagus* species were monoterpene emitters (Table 2), whereas they did not emit a detectable amount of isoprene. The three *Nothofagus* species had also inherently different monoterpene emission rates, but that was not related to the characteristic of deciduousness of three species (Table 1). The two deciduous species *N. pumilio* and *N. antarctica* had similar photosynthetic parameters, but monoterpene emission rate was more than three-fold, and the fraction of photosynthetic carbon lost was two-fold higher in *N. pumilio* than in *N. Antarctica*. The evergreen species *N. dombeyi* showed intermediate values for total monoterpene emissions, but highest values for the fraction of photosynthetic carbon lost. The emission rates of monoterpenes in the three *Nothofagus* species were similar to those observed in other monoterpene emitting species of the *Fagaceae* family, all of which belong to the *Quercus* genus (Loreto et al. 1998; Loreto 2002).

There is a debate on whether the *Nothofagus* genus, the only genus of the *Fagaceae* family in the southern hemisphere, is closely linked to plants of the family living in the northern hemisphere (Manos and Steele 1997; Li et al. 2004). Previous studies considered the *Nothofagus* family as genetically close to the *Fagus* genus (Cronquist 1981). However, Manos and Steele (1997), using phylogenetic analyses based on chloroplast DNA sequences, suggested that *Nothofagus* is largely independent genus, constituting a clade different than that including other *Fagaceae* (Manos and Steele 1997). More recent studies confirmed that *Nothofagus* is monophyletic and sister to the rest of *Fagales* (Li et al. 2004; Cook and Crisp 2005).

Isoprenoids have been used as a trait for systematic studies (Loreto et al. 1998; Lerdau and Gray 2003). Fagus sylvatica has been considered as a 'nonemitting' or low monoterpene emitting species (Hewitt and Street 1992; König et al. 1995), but several recent experiments in field conditions have shown that the emission of monoterpenes may be more relevant than previously indicated (Kahl et al. 1999; Kesselmeier and Staudt 1999; Dindorf et al. 2006; Holzke et al. 2006), although probably still lower than that reported in strong emitters such as some Mediterranean oak species (Loreto 2002). The rates of monoterpene emission by leaves of the three *Nothofagus* species were quite high, similar to those of Mediterranean oaks. However, the blend emitted from Nothofagus plants was different than those emitted by Fagaceae (Figure 1). In fact, sabinene and α -thuyene are the predominant compounds released by Fagus sylvatica leaves (Dindorf et al. 2006; Holzke et al. 2006), while α -pinene is the main

Table 1. Photosynthesis (A), stomatal conductance (g_s), respiration in darkness (R_D), total monoterpene emission, and percentage of photosynthetic carbon lost as isoprenoids in *Nothofagus antarctica* (deciduous), *N. pumilio* (deciduous) and *N. dombeyi* (evergreen). Data are means of 12–15 trees ±1 SEM. Letters (a, b, c) indicate significant differences at p < 0.05 in the same column.

	$A \ (\mu mol \ m^{-2}s^{-1})$	$g_{\rm s} ({\rm mmol}{\rm m}^{-2}{\rm s}^{-1})$	$R_{\rm D} (\mu { m mol} { m m}^{-2}{ m s}^{-1})$	Monoterpene $(nmol m^{-2}s^{-1})$	% Carbon lost
N. antarctica	6.02 ± 0.76 b	76.00 ± 13.87 b	-2.93 ± 0.32 b	2.63 ± 0.31 a	0.141 a
N. pumilio	5.93 ± 0.82 b	90.00 ± 7.71 b	-2.90 ± 0.23 b	8.02 ± 1.30 c	0.342 b
N. dombeyi	4.14 ± 0.53 a	34.83 ± 3.97 a	-1.77 ± 0.06 a	4.38 ± 1.20 b	0.437 c

Table 2. Emission of the most abundant monoterpenes (nmol m⁻²s⁻¹) in *Nothofagus antarctica* (deciduous), *N. pumilio* (deciduous) and *N. dombeyi* (evergreen). Data are means of 12–15 trees ± 1 SEM. Letters (a, b, c) indicate significant differences at p < 0.05 in the same line.

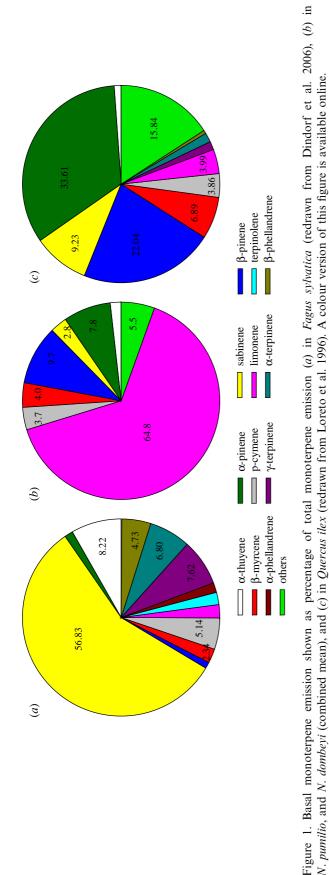
Monoterpene	N. antarctica	N. pumilio	N. dombeyi
α-thuyene	0.040±0.011 a	0.233±0.045 b	0.039 ± 0.006 a
α-pinene	0.241 ± 0.061 a	0.620±0.080 b	0.278±0.087 a
sabinene	0.077 ± 0.012 a	0.163±0.050 b	0.149±0.045 b
β-pinene	0.325 ± 0.050 a	0.540±0.082 b	0.441 ± 0.072 ab
β-myrcene	0.124 ± 0.027 a	0.348±0.073 b	0.137 ± 0.030 a
p-cymene	0.070 ± 0.011 a	0.362 ± 0.098 c	0.165±0.042 b
limonene	1.607 ± 0.175 a	5.332 ± 1.069 c	2.920 ± 0.690 b

compound emitted by Quercus ilex leaves (Loreto et al. 1996). In all Nothofagus species limonene was found to be the most abundant monoterpene emitted (Table 2), accounting for about 61-67% of the total emission, followed by β -pinene and α -pinene (Figure 1). This is interpreted as a possible indication that Nothofagus is genetically different from both Quercus and Fagus plant species, thus confirming recent phylogenetic analysis (Manos and Steele 1997). It should be mentioned that limonene has been reported as the main component of the blend emitted by some provenances of Quercus suber, Quercus ilex and hybrids of these two species (Staudt et al. 2004). It may be possible that the emission of limonene is a trait under environmental control (Geron et al. 2000). This should be further investigated under controlled environmental conditions.

Only evergreen species of the *Quercus* family of the northern hemisphere emit monoterpenes, while deciduous species only emit isoprene (Loreto 2002). A remarkable exception appears to be *Q. semecarpifolia*, an Asian species that only emit isoprene (Loreto et al. 1998). It is therefore noteworthy that the spectrum of the main monoterpene emitted is very similar among the two deciduous and the evergreen *Nothofagus* species, indicating the absence of differentiation based on the persistence of the leaf. This is another observation that may be explained by the environmental conditions experienced by the plants, as they share the same habitat and might have evolved the same emission pattern, independent of deciduousness.

The similar emission pattern in the three Nothofagus species might also indicate a substantially low interspecific difference at a genetic level. Manos (1997) examined the phylogenetic relationships within the Nothofagaceae family by analyzing the nucleotide sequences and their morphological character evolution from 22 Nothofagus species representing the four currently recognized subgenera (i.e. Brassospora, Nothofagus, Fuscospora and Lophozonia) and related outgroups. He showed that N. pumilio, N. antarctica and N. dombeyi belonged to the subgenus Nothofagus. Furthermore, Ramírez et al. (1997), using a morphological criterion, divided 11 taxa of South American Nothofagus in five groups. They found that N. antarctica and N. dombeyi belong to the same group having small leaves and that developed under temperate humid conditions; whereas N. pumilio, with mid-sized deciduous leaves, adapted to the cold and dry zones of the southern Andes, belongs to a group of isolated species with different requirements. However, N. pumilio and the group including *N. antarctica* and *N. dombeyi* all derived directly from the most primitive group formed by the Nothofagus species with large deciduous leaves (i.e. N. obliqua, N. obliqua var. macrocarpa, N. leoni and N. alessandrii). Recently, Stecconi et al. (2004) compared N. antarctica and N. dombeyi with their putative hybrid individuals found in natural stands by using morphological and isoenzymatic traits, and pointed out that N. antarctica and N. dombeyi are probably more closely related than previously assumed. Thus, the similarity in the monoterpene emission spectrum among the three screened species may mirror this common origin. Because the Nothofagus genus (formed by 40 species) has a distributional range that comprises the southern end of South America, Australia, New Zealand, New Caledonia, New Guinea and other small Pacific islands, the monoterpene emission spectrum may be a useful 'tool' to ascertain the phytogeographic origin and the phytogenic evolution of the Nothofagus genus.

Guenther et al. (2006) and Geron et al. (2006) have recently produced regional isoprenoid emission inventories, and have shown that species-level isoprenoid emission data are useful for assessing the potential impacts on air quality at regional and global scale. Geron et al. (2006) pointed out that the dramatic landcover changes occurring in South Asia, such as increasing in plantation area established with high isoprene emitting species (e.g. Bambusa spp. and Eucalyptus spp.), can lead to increases in BVOC emissions in the near future, and this may potentially increase the formation of tropospheric ozone altering the air quality over this rapidly developing region. The Nothofagus species dominate the temperate forests present in southern Chile and Argentina, which consist of evergreen, deciduous, needle and broadleaf forests occurring at latitudes south of the 30° S parallel (Veblen et al. 1996). This favors monoterpene emission over isoprene, and



antarctica.

N.

may influence the biogenic SOA produced in this region, despite the *Nothofagus* species are low emitters of sabinene (Table 2, Figure 1), which is the compound with higher new particle formation rate compared to other common monoterpenes (Koch et al. 2000). However, rapid industrial development and landcover changes may significantly affect future isoprenoid fluxes in southern Chile and Argentina. Thus, further inventory studies and advanced modeling system are needed to assess BVOC emissions and their impact on air quality and atmospheric chemistry in this region.

In conclusion, the experiments performed in the present study clearly indicate, for the first time to our knowledge, that Nothofagus spp. emits large quantities of monoterpenes, with limonene being the predominant compound released. We believe that this finding may be useful for a better parameterization of large scale emission models, because the current predictions on the impact of isoprenoid emissions on air quality at regional and global scale is currently limited by the available global emission inventories which still lack information from a large number of unmeasured species (Grote and Niinemets in press), especially from South American vegetation other than the rainforest ecosystem. Moreover, the three Nothofagus species tested had inherently different photosynthetic traits that were related to the characteristic of deciduousness of three species, whereas monoterpene emission rates were surprisingly independent from this character, an observation only rarely replicated worldwide. Finally, the monoterpene emission spectrum is very similar among the three Nothofagus species screened. This trait, that may have an important phytogeographic and phytogenic relevance, may spring from the same common origin.

Acknowledgements

This research was financially supported by the Argentinean SECyT and Italian MAE-Programme of Scientific and Technological Cooperation 2004–2005 (IT/PA03 – BVII/072), and by the Argentinean Instituto de Clima y Agua (INTA) Projects AERN4 and AERN4642.

References

- Andreae MO, Crutzen PJ. 1997. Atmospheric aerosols: Biogeochemical sources and role in atmospheric chemistry. Science. 276:1052–1058.
- Atkinson R, Arey J. 2003. Gas-phase tropospheric chemistry of biogenic volatile organic compounds: a review. Atmos Environ. 37:197–219.
- Baraldi R, Rapparini F, Rossi F, Latella A, Ciccioli P. 1999. Volatile organic compound emissions from flowers of the most occurring and economically important fruit tree species. Phys Chem Earth 6:729– 732.
- Brilli F, Barta C, Fortunati A, Lerdau M, Loreto F, Centritto M. 2007. Response of isoprene emission

and carbon metabolism to drought in white poplar saplings. New Phytol. 175:244–254.

- Centritto M, Nascetti P, Petrilli L, Raschi A, Loreto F. 2004. Profiles of isoprene emission and photosynthetic parameters in hybrid poplars exposed to free-air CO₂ enrichment. Plant Cell Environ. 27:403–412.
- Chameides W, Lindsay R, Richardson J, et al. 1987. The role of biogenic hydrocarbons in urban photochemical smog: Atlanta as a case study. Science. 241:1473–1475.
- Cook LG, Crisp MD. 2005. Not so ancient: the extant crown group of *Nothofagus* represents a post-Gondwanan radiation. Proc R Soc B. 272:2535–2544.
- Cronquist A. 1981. An integrated system of classification of flowering plants. New York (NY): Columbia University Press.
- Di Carlo P, Brune WH, Martinez M, Harder H, Lesher R, Ren X, Thornberry T, Carroll MA, Young V, Shepson PB, Riemer D, Apel E, Campbell C. 2004. Missing OH reactivity in a forest: evidence for unknown reactive biogenic VOCs. Science. 304:722–725.
- Dindorf T, Kuhn U, Ganzeveld L, Schebeske G, Ciccioli P, Holzke C, Köble R, Seufert G, Kesselmeier J. 2006. Significant light and temperature dependent monoterpene emissions from European beech (*Fagus sylvatica* L.) and their potential impact on the European volatile organic compound budget. J Geophys Res. 111, D16305, DOI: 10.1029/2005JD006751.
- Fehsenfeld F, Calvert J, Fall R, Goldan P, Guenther A, Hewitt C, Lamb B, Liu S, Trainer M, Westberg H, Zimmerman P. 1992. Emissions of volatile organic compounds from vegetation and the implications for atmospheric chemistry. Global Biochem Cycles. 6:389– 430.
- Geron C, Owen S, Guenther A, Greenberg J, Rasmussen R, Bai JH, Li Q-J, Baker B. 2006. Volatile organic compounds from vegetation in southern Yunnan Province, China: emission rates and some potential regional implications. Atmos Environ. 40:1759–1773.
- Geron C, Rasmussen R, Arnts RR, Guenther A. 2000. A review and synthesis of monoterpene speciation. Atmos Environ. 34:1761–1781.
- Gershenzon J, Dudareva N. 2007. The function of terpene natural products in the natural world. Nature Chem Biol. 3:408–414.
- Greenberg JP, Guenther AB, Pètron G, Wiedinmyer C, Vega O, Gatti LV, Tota J, Fisch G. 2004. Biogenic VOC emissions from forested Amazonian landscapes. Global Change Biol. 10:651–662.
- Grote R, Niinemets Ü. In press. Modeling volatile isoprenoid emissions – a story with split ends. Plant Biol. DOI: 10.1055/s-2007-964975.
- Guenther A, Geron C, Pierce T. 2000. Natural emissions of non-methane volatile organic compounds, carbon monoxide, and oxides of nitrogen from North America. Atmos Environ. 34:2205–2230.
- Guenther A, Greenberg J, Harley P, Helmig D, Klinger L, Vierling L, Zimmerman P, Geron C. 1996a. Leaf, branch, stand and landscape scale measurements of volatile organic compound fluxes from U.S. woodlands. Tree Physiol. 16:17–24.
- Guenther A, Karl T, Harley P, Wiedinmyer C, Palmer PI, Geron C. 2006. Estimates of global terrestrial isoprene emissions using MEGAN (Model of Emissions of Gases and Aerosols from Nature). Atmos Chem. Phys. 6:3181–3210.

- Guenther A, Otter L, Zimmerman P, Greenberg J, Scholes R, Scholes M. 1996b. Biogenic hydrocarbon emissions from southern African savannas. J Geophys Res. 101:25859–25865.
- Guenther A, Zimmermann P, Harley PC, Monson RK, Fall
 R. 1993. Isoprene and monoterpene emission rate variability: model evaluations and sensitivity analysis.
 J Geophys Res. 98:12609–12617.
- He C, Murray F, Lyons T. 2000. Monoterpene and isoprene emissions from 15 *Eucalyptus* species in Australia. Atmos Environ. 34:645–655.
- Hewitt CN, Street RA. 1992. A qualitative assessment of the emission of non-methane hydrocarbon compounds from the bisophere to the atmosphere in the UK: present knowledge and uncertainties. Atmos Environ. 26:3069–3077.
- Holzke C, Dindorf T, Kesselmeier J, Kuhn U, Koppmann R. 2006. Terpene emissions from European beech (*Fagus sylvatica* L.): pattern and emission behaviour over two vegetation periods. J Atmos Chem. 55:81– 102.
- Kahl J, Hoffmann T, Klockow D. 1999. Differentiation between de novo synthesized and constitutively released terpenoids from *Fagus sylvatica*. Phytochemistry. 51:383–388.
- Kanakidou M, Seinfeld JH, Pandis SN, Barnes I, Dentener FJ, Facchini MC, van Dingenen R, Ervens B, Nenes A, Nielsen CJ, et al. 2004. Organic aerosol and global climate modelling: a review. Atmos Chem Phys. Discuss. 4:5855–6024.
- Kavouras IG, Mihalopoulos N, Stephanou EG. 1998. Formation of atmospheric particles from organic acids produced by forests. Nature. 395:683–686.
- Kesselmeier J, Staudt M. 1999. Biogenic volatile organic compounds (VOC): an overview on emission, physiology and ecology. J Atmos Chem. 33:23–88.
- Koch S, Winterhalter R, Uherek E, Kolloff A, Neeb P, Moortgat GK. 2000. Formation of new particles in the gasphase ozonolysis of monoterpenes. Atmos Environ. 34:4031–4042.
- König G, Brunda M, Puxbaum H, Hewitt CN, Duckham SC, Rudolph J. 1995. Relative contribution of oxygenated hydrocarbons to the total biogenic VOC emissions of selected mid-European agricultural and natural plant species. Atmos Environ. 29:861–874.
- Lal R. 1999. World soils and the greenhouse effect. IGBP Newsl. 37:4–5.
- Lerdau M, Gray D. 2003. Ecology and evolution of lightdependent and light-independent phytogenic volatile organic carbon. New Phytol. 157:199–211.
- Li RQ, Chen ZD, Lu AM, Soltis DE, Soltis PS, Manos PS. 2004. Phylogenetic relationships in *Fagales* based on DNA sequences from three genomes. Int J Plant Sci. 165:311–324.
- Loreto F. 2002. Distribution of isoprenoid emitters in the *Quercus* genus around the world: chemo-taxonomical implications and evolutionary considerations based on the ecological function of the trait. Perspec Plant Eco Evol Syst. 5:185–192.
- Loreto F, Centritto M, Baraldi R, Rapparini F, Liu S. 2002. Emission of isoprenoids from natural vegetation in the Beijing region (Northern China). Plant Biosyst. 136:251–256.
- Loreto F, Ciccioli P, Brancaleoni E, Cecinato A, Frattoni M, Sharkey TD. 1996. Different sources of reduced

carbon contribute to form three classes of terpenoid emitted by *Quercus ilex* L. leaves. Proc Natl Acad Sci. 93:9966–9969.

- Loreto F, Ciccioli P, Brancaleoni E, Valentini R, De Lillis M, Csiky O, Seufert G. 1998. A hypothesis on the evolution of isoprenoid emission by oaks based on the correlation between emission type and *Quercus* taxonomy. Oecologia 115:302–305.
- Manos PS. 1997. Systematics of Nothofagus (*Nothofaga-ceae*) based on rDNA spacer sequences (ITS): taxo-nomic congruence with morphology and plastid sequences. Am J Bot. 84:1137–1155.
- Manos PS, Steele KP. 1997. Phylogenetic analyses of 'higher' hamamelididae based on plastid sequence data. Am J Bot. 84:1407–1419.
- Niinemets Ü, Hauff K, Bertin N, Tenhunen JD, Steinbrecher R, Seufert G. 2002. Monoterpene emissions in relation to foliar photosynthetic and structural variables in Mediterranean evergreen *Quercus* species. New Phytol. 153:243–256.
- Niinemets Ü, Loreto F, Reichstein M. 2004. Physiological and physico-chemical controls on foliar volatile organic compound emissions. Trends Plant Sci. 9:180– 186.
- Poisson N, Kanakidou M, Crutzen PJ. 2000. Impact of non-methane hydrocarbons on tropospheric chemistry and the oxidizing power of the global troposphere: 3dimensional modelling results. J Atmos Chem. 36:157– 230.
- Ramírez C, San Martín C, Oyarzún A, Figueroa H. 1997. Morpho-ecological study on the South American species of the genus *Nothofagus*. Plant Ecol. 130:101– 109.
- Rasmussen RA, Jones CA. 1973. Emission isoprene from leaf discs of *Hamamelis*. Phytochemistry. 12:15–19.

- Reich PB, Walters MB, Ellesworth DS. 1997. From tropics to tundra: Global convergence in plant functioning. Proc Natl Acad Sci USA. 94:13730–13734.
- Rinne HJI, Guenther AB, Greenberg JP, Harley PC. 2002. Isoprene and monoterpene fluxes measured above Amazonian rainforest and their dependence on light and temperature. Atmos Environ. 36:2421–2426.
- Sharkey TD, Yeh S. 2001. Isoprene emission from plants. Ann Rev Plant Physiol Plant Mol Biol. 52:407–436.
- Simpson D, Guenther A, Hewitt CN, Steinbrecher R. 1995. Biogenic emissions in Europe 1. Estimates and uncertainties. J Geophys Res. 100:22875–22890.
- Staudt M, Mir C, Joffre R, Rambal S, Bonin A, Landais D, Lumaret R. 2004. Isoprenoid emissions of *Quercus* spp. (*Q. suber* and *Q. ilex*) in mixed stands contrasting in interspecific genetic introgression. New Phytol. 163:573–584.
- Stecconi M, Marchelli P, Puntieri J, Picca PI, Gallo L. 2004. Natural hybridization between a deciduous (*Nothofagus antarctica*, *Nothofagaceae*) and an evergreen (*N. dombeyi*) forest tree species: evidence from morphological and isoenzymatic traits. Ann Bot. 94:775–786.
- Veblen TT, Donoso C, Kitzberger T, Rebertus AJ. 1996.
 Ecology of Southern Chilean and Argentinean *Notho-fagus* forests. In: Veblen TT, Hill RS, Read J, editors.
 The ecology and biogeography of *Nothofagus* forests.
 New Haven: Yale University Press. p 293–353.
- Vizuete W, Junquera V, Allen DT. 2004. Sesquiterpene emissions and secondary organic aerosol formation potentials for southeast Texas. Aerosol Sci Technol. 38:167–181.
- Wright IJ, Groom PK, Lamont BB, Poot P, Prior LD, Reich PB, Schulze E-D, Veneklaas E, Westoby M. 2004. Leaf trait relationships in Australian plant species. Funct Plant Biol. 31:551–558.

Copyright of Journal of Plant Interactions is the property of Taylor & Francis Ltd and its content may not be copied or emailed to multiple sites or posted to a listserv without the copyright holder's express written permission. However, users may print, download, or email articles for individual use.